PCT

WORLD INTELLECTUAL PROPERTY ORGANIZATION
International Bureau

INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification 5 :
A01N 63/00, 63/02, C12N 1/20
// (C12N 1/20, C12R 1:07)

A1

(11) International Publication Number: WO 92/19106

(43) International Publication Date: 12 November 1992 (12.11.92)

(21) International Application Number: PCT/US92/03546

(22) International Filing Date: 30 April 1992 (30.04.92)

(30) Priority data:

693,210	30 April 1991 (30.04.91)	US
759,248	13 September 1991 (13.09.91)	US
768,141	30 September 1991 (30.09.91)	US

(71) Applicant: MYCOGEN CORPORATION [US/US]; 5451 Oberlin Drive, San Diego, CA 92121 (US).

(72) Inventors: PAYNE, Jewel, M. ; 7984 Hemphill Drive, San Diego, CA 92126 (US). CANNON, Raymond, J., C. ; 6 Barn Close, Borden, Sittingbourne, Kent ME9 8JW (GB). BAGLEY, Angela, L. ; Lion Farm Cottage, Milstead, Sittingbourne, Kent ME9 0RT Centre (GB).

(74) Agents: SALIWANCHIK, David, R. et al.; Saliwanchik & Saliwanchik, 2421 N.W. 41st Street, Suite A-1, Gainesville, FL 32606 (US).

(81) Designated States: AT (European patent), AU, BE (European patent), CA, CH (European patent), DE (European patent), DK (European patent), ES (European patent), FR (European patent), GB (European patent), GR (European patent), IT (European patent), JP, KR, LU (European patent), MC (European patent), NL (European patent), RU, SE (European patent).

Published
With international search report.
Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.

(54) Title: NOVEL *BACILLUS THURINGIENSIS* ISOLATES FOR CONTROLLING ACARIDES

(57) Abstract

Disclosed and claimed are *Bacillus thuringiensis* isolates designated *B.t.* PS50C, *B.t.* PS86A1, *B.t.* PS69D1, *B.t.* PS72L1, *B.t.* PS75J1, *B.t.* PS83E5, *B.t.* PS45B1, *B.t.* PS24J, *B.t.* PS94R3, *B.t.* PS17, *B.t.* PS62B1 and *B.t.* PS74G1 which are active against acaride pests. Thus, these isolates, or mutants thereof, can be used to control such pests. Further, genes encoding novel δ-endotoxins can be removed from these isolates and transferred to other host microbes, or plants. Expression of the δ-endotoxins in microbe hosts results in the control of acaride pests, whereas transformed plants become resistant to acaride pests.

FOR THE PURPOSES OF INFORMATION ONLY

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AT	Austria	ES	Spain	MG	Madagascar
AU	Australia	FI	Finland	ML	Mali
BB	Barbados	FR	France	MN	Mongolia
BE	Belgium	GA	Gabon	MR	Mauritania
BF	Burkina Faso	GB	United Kingdom	MW	Malawi
BG	Bulgaria	GN	Guinea	NL	Netherlands
BJ	Benin	GR	Greece	NO	Norway
BR	Brazil	HU	Hungary	PL	Poland
CA	Canada	IT	Italy	RO	Romania
CF	Central African Republic	JP	Japan	RU	Russian Federation
CG	Congo	KP	Democratic People's Republic of Korea	SD	Sudan
CH	Switzerland			SE	Sweden
CI	Côte d'Ivoire	KR	Republic of Korea	SN	Senegal
CM	Cameroon	LI	Liechtenstein	SU	Soviet Union
CS	Czechoslovakia	LK	Sri Lanka	TD	Chad
DE	Germany	LU	Luxembourg	TG	Togo
DK	Denmark	MC	Monaco	US	United States of America

DESCRIPTION

NOVEL *BACILLUS THURINGIENSIS* ISOLATES FOR CONTROLLING ACARIDES

Cross-Reference to a Related Application

This is a continuation-in-part of co-pending application Serial No. 07/693,210, filed on April 30, 1991. This is also a continuation-in-part of application Serial No. 07/768,141, filed on September 30, 1991 which is a continuation-in-part of application Serial No. 07/759,248, filed on September 13, 1991.

Background of the Invention

The spore-forming microorganism Bacillus thuringiensis (B.t.) produces the best-known insect toxin. The toxin is a protein, designated as δ-endotoxin. It is synthesized by the B.t. sporulating cell. The toxin, upon being ingested in its crystalline form by susceptible insect larvae, is transformed into biologically active moieties by the insect gut juice proteases. The primary target is insect cells of the gut epithelium, which are rapidly destroyed. Experience has shown that the activity of the B.t. toxin is so high that only nanogram amounts are required to kill susceptible insects.

The reported activity spectrum of B.t. covers insect species within the order Lepidoptera, which is a major insect problem in agriculture and forestry. The activity spectrum also includes the insect order Diptera, wherein reside mosquitoes and blackflies. See Couch, T.L., (1980) "Mosquito Pathogenicity of Bacillus thuringiensis var. israelensis," Developments in Industrial Microbiology, 22:61-67; Beegle, C.C, (1978) "Use of Entomogeneous Bacteria in Agroecosystems," Developments in Industrial Microbiology, 20:97-104.

U.S. Patent 4,771,131 discloses a toxin gene isolated from a strain of Bacillus thuringiensis. This gene encodes a toxin which is active against beetles of the order Coleoptera.

There have been published reports concerning the use of Bacillus thuringiensis preparations for the control of mites. These publications are as follow:

Royalty, R.N., Hall, F.R. and Taylor, R.A.J. 1990. Effects of thuringiensin on Tetranychus urticae (Acari: Tetranychidae) mortality, fecundity, and feeding. J. Econ. Entomol. 83:792-798.

Neal, J.W., Lindquist, R.K., Gott, K.M. and Casey, M.L. 1987. Activity of the themostable beta-exotoxin of Bacillus thuringiensis Berliner on Tetranychus urticae and Tetranychus cinnabarinus. J. Agric. Entomol. 4:33-40.

Vlayen, P., Impe, G. and Van Semaille, R. 1978. Effect of a commercial preparation of Bacillus thuringiensis on the spider mite Tetranychus urticae Koch. (Acari: Tetranychidae). Mededelingen 43:471-479.

In the above published studies, the active ingredient in the B.t. preparations was beta-exotoxin (also called thuringiensin).

U.S. Patent No. 4,695,455 concerns methods and compositions for preparing and using biological pesticides, where the pesticides are encapsulated in non-proliferating cells.

U.S. Patent No. 4,849,217 concerns B.t. isolates active against the alfalfa weevil.

Brief Summary of the Invention

The subject invention concerns Bacillus thuringiensis isolates and toxins which have acaricidal properties. Unlike published reports of the use of B.t. ß-exotoxins to control mites, the subject invention isolates express δ-endotoxins which control mites. The use of δ-endotoxins is highly advantageous in view of the known general toxicity of ß-exotoxins to humans and animals.

More specifically, the subject invention concerns Bacillus thuringiensis isolates designated B.t. PS50C, B.t. PS86A1, B.t. PS69D1, B.t. PS72L1, B.t. PS75J1, B.t. PS83E5, B.t. PS45B1, B.t. PS24J, B.t. PS94R3, B.t. PS17, B.t. PS62B1 and B.t. PS74G1.

The B.t. isolates of the subject invention are toxic to the Two Spotted Spider Mite, Tetranychus urticae. Thus, these isolates can be used to control this mite. Further, the δ-endotoxins from these B.t. isolates can be isolated by standard procedures, e.g. ion exchange, and formulated by standard procedures to control the Two Spotted Spider Mite. These B.t. isolates can also be used against non-phytophagus mites such as acarid pests of livestock, fowl and stored products. Still further, the gene(s) from the B.t. isolates of the invention which encode the acaricidal toxin can be cloned from the isolates and then used to transform other hosts, e.g., prokaryotic, eukaryotic or plants, which transformed host can be used to control mites, or, in the case of transgenic plants, be resistant to mites.

Brief Description of the Drawings

FIGURES 1, 2A and 2B are photographs of 12% SDS polyacrylamide gels showing alkali-soluble proteins of the isolates of the invention.

Brief Description of the Sequences

SEQ ID NO. 1 discloses the DNA of 17a.

SEQ ID NO. 2 discloses the amino acid sequence of the toxin encoded by 17a.

SEQ ID NO. 3 discloses the DNA of 17b.

SEQ ID NO. 4 discloses the amino acid sequence of the toxin encoded by 17b.

SEQ ID NO. 5 is the nucleotide sequence of gene 33F2.

SEQ ID NO. 6 is the nucleotide sequence f a gene from 52A1.

SEQ ID NO. 7 is the amino acid sequence of the protein expressed by the gene from 52A1.

SEQ ID NO. 8 is the nucleotide sequence of a gene from 69D1.

SEQ ID NO. 9 is the amino acid sequence of the protein expressed by the gene from 69D1.

SEQ ID NO. 10 is the DNA coding for the amino acid sequence of SEQ ID NO. 13.

SEQ ID NO. 11 is the amino acid sequence of a probe which can be used according to the subject invention.

SEQ ID NO. 12 is the N-terminal amino acid sequence of 17a.

SEQ ID NO. 13 is the N-terminal amino acid sequence of 17b.

SEQ ID NO. 14 is the N-terminal amino acid sequence of 52A1.

SEQ ID NO. 15 is the N-terminal amino acid sequence of 69D1.

SEQ ID NO. 16 is a synthetic oligonucleotide derived from 17.

SEQ ID NO. 17 is an oligonucleotide probe designed from the N-terminal amino acid sequence of 52A1.

SEQ ID NO. 18 is the synthetic oligonucleotide probe designated as 69D1-D.

SEQ ID NO. 19 is the forward oligonucleotide primer from 63B.

SEQ ID NO. 20 is the reverse complement primer to SEQ ID NO. 29, used according to the subject invention.

SEQ ID NO. 21 is the DNA coding for the primer of SEQ ID NO. 31.

SEQ ID NO. 22 is a forward primer according to the subject invention.

SEQ ID NO. 23 is a probe according to the subject invention.

SEQ ID NO. 24 is a probe according to the subject invention.

SEQ ID NO. 25 is a probe according to the subject invention.

SEQ ID NO. 26 is a forward primer according to the subject invention.

SEQ ID NO. 27 is the nucleotide sequence of a gene from PS50C.

SEQ ID NO. 28 is the amino acid sequence of the protein expressed by the gene from PS50C.

SEQ ID NO. 29 is the nucleotide sequence of a gene from PS86A1.

SEQ ID NO. 30 is the amino acid sequence of the protein expressed by the gene from PS86A1.

Detailed Disclosure of the Invention

The subject invention concerns B.t. δ-endotoxins having acaricidal activity. In addition to having acaricidal activity, the toxins of the subject invention may have one or more of the following characteristics:

1. A high degree of amino acid homology with specific toxins disclosed herein.
2. A DNA sequence encoding the toxin which hybridizes with probes or genes disclosed herein.
3. A nucleotide sequence which can be amplified using primers disclosed herein.
4. Immunoreactivity to an antibody raised to a specific toxin disclosed herein.

Acaride-active toxins according to the subject invention are specifically exemplified herein by the toxins encoded by the genes designated 17a, 17b, and 69D1. Since these toxins are merely exemplary of the toxins presented herein, it should be readily apparent that the subject invention further comprises toxins from the other disclosed isolates as well as equivalent toxins (and nucleotide sequences coding for equivalent toxins) having the same or similar biological activity of the specific toxins disclosed or claimed herein. These equivalent toxins will have amino acid homology with the toxins disclosed and claimed herein. This amino acid homology will typically be greater than 50%, preferably be greater than 75%, and most preferably be greater than 90%. The amino acid homology will be highest in certain critical regions of the toxin which account for biological activity or are involved in the determination of three-dimensional configuration which ultimately is responsible for the biological activity. In this regard, certain amino acid substitutions are acceptable and can be expected if these substitutions are in regions which are not critical to activity or are conservative amino acid substitutions which do not affect the three-dimensional configuration of the molecule. For example, amino acids may be placed in the following classes: non-polar, uncharged polar, basic, and acidic. Conservative substitutions whereby an amino acid of one class is replaced with another amino acid of the same type fall within the scope of the subject invention so long as the substitution does not materially alter the biological activity of the compound. Table 1 provides a listing of examples of amino acids belonging to each class.

Table 1

Class of Amino Acid	Examples of Amino Acids
Nonpolar	Ala, Val, Leu, Ile, Pro, Met, Phe, Trp
Uncharged Polar	Gly, Ser, Thr, Cys, Tyr, Asn, Gln
Acidic	Asp, Glu
Basic	Lys, Arg, His

In some instances, non-conservative substitutions can also be made. The critical factor is that these substitutions must not significantly detract from the biological activity of

the toxin. The information presented in the generic formulae of the subject invention provides clear guidance to the person skilled in this art in making various amino acid substitutions.

The B.t. isolates of the invention have the following characteristics:

Strain		Crystal Type	Approx. Mol. Wt. of Proteins (kDa)
B. thuringiensis	PS50C	Sphere	135 doublet
B. thuringiensis	PS86A1	Multiple	45, 58
B. thuringiensis	PS69D1	Elongated	34, 48, 145
B. thuringiensis	PS72L1	Long rectangle	42, 50
B. thuringiensis	PS75J1	Amorphic	63, 74, 78, 84
B. thuringiensis	PS83E5	Multiple	37, 42
B. thuringiensis	PS24J	Long	51, 48, 43
B. thuringiensis	PS94R3	Long	50, 43, 42
B. thuringiensis	PS45B1	Multiple	150, 135, 35
B. thuringiensis	PS17	Long	155, 145, 128
B. thuringiensis	PS62B1	Attached multiple	35
B. thuringiensis	PS74G1	Amorphic	148, 112, 104, 61

Additionally, the isolates have the following common characteristics:

Colony morphology -- large colony, dull surface, typical B.t.

Vegetative cell morphology -- typical B.t.

The toxins of the subject invention can be accurately characterized in terms of the shape and location of crystal toxin inclusions. Specifically, acaride-active inclusions typically remain attached to the spore after cell lysis. These inclusions are not inside the exosporium, as in previous descriptions of attached inclusions, but are held within the spore by another mechanism. Inclusions of the acaride-active isolates are typically amorphic, generally long and/or multiple. These inclusions are distinguishable from the larger round/amorphic inclusions that remain attached to the spore. No B.t. strains that fit this description have been found to have activity against the conventional targets—Lepidoptera, Diptera, or Colorado Potato Beetle. We have found a very high correlation between this crystal structure and acaride activity.

The genes and toxins according to the subject invention include not only the full length sequences disclosed herein but also fragments of these sequences, or fusion proteins, which retain the characteristic acaricidal activity of the sequences specifically exemplified herein.

It should be apparent to a person skilled in this art that genes coding for acaride-active toxins can be identified and obtained through several means. The specific genes may be obtained from a culture depository as described below. These genes, or portions thereof, may be constructed synthetically, for example, by use of a gene machine. Variations of these genes may be readily constructed using standard techniques for making point mutations. Also, fragments of these genes can be made using commercially available exonucleases or endonucleases according to standard procedures. For example, enzymes such as Bal31 or site-directed mutagenesis can be used to systematically cut ff nucleotides from the ends of these

genes. Also, genes which code for active fragments may be obtained using a variety of other restriction enzymes. Proteases may be used to directly obtain active fragments of these toxins.

Equivalent toxins and/or genes encoding these equivalent toxins can also be located from B.t. isolates and/or DNA libraries using the teachings provided herein. There are a number of methods for obtaining the acaride-active toxins of the instant invention which occur in nature. For example, antibodies to the acaride-active toxins disclosed and claimed herein can be used to identify and isolate other toxins from a mixture of proteins. Specifically, antibodies may be raised to the acaride-active toxins using procedures which are well known in the art. These antibodies can then be used to specifically identify equivalent toxins with the characteristic acaricidal activity by immunoprecipitation, enzyme linked immunoassay (ELISA), or Western blotting. Antibodies to the toxins disclosed herein, or to equivalent toxins, or fragments of these toxins, can readily be prepared using standard procedures in this art. The genes coding for these toxins can then be obtained from the microorganism.

A further method for identifying the toxins and genes of the subject invention is through the use of oligonucleotide probes. These probes are nucleotide sequences having a detectable label. As is well known in the art, if the probe molecule and nucleic acid sample hybridize by forming a strong bond between the two molecules, it can be reasonably assumed that the probe and sample are essentially identical. The probe's detectable label provides a means for determining in a known manner whether hybridization has occurred. Such a probe analysis provides a rapid method for identifying nematicidal endotoxin genes of the subject invention.

The nucleotide segments which are used as probes according to the invention can be synthesized by use of DNA synthesizers using standard procedures. In the use of the nucleotide segments as probes, the particular probe is labeled with any suitable label known to those skilled in the art, including radioactive and non-radioactive labels. Typical radioactive labels include ^{32}P, ^{125}I, ^{35}S, or the like. A probe labeled with a radioactive isotope can be constructed from a nucleotide sequence complementary to the DNA sample by a conventional nick translation reaction, using a DNase and DNA polymerase. The probe and sample can then be combined in a hybridization buffer solution and held at an appropriate temperature until annealing occurs. Thereafter, the membrane is washed free of extraneous materials, leaving the sample and bound probe molecules typically detected and quantified by autoradiography and/or liquid scintillation counting.

Non-radioactive labels include, for example, ligands such as biotin or thyroxine, as well as enzymes such as hydrolases or perixodases, or the various chemiluminescers such as luciferin, or fluorescent compounds like fluorescein and its derivatives. The probe may also be labeled at both ends with different types of labels for ease of separation, as, for example, by using an isotopic label at the end mentioned above and a biotin label at the ther end.

Duplex formation and stability depend on substantial complementarity between the two strands of a hybrid, and, as noted above, a certain degree of mismatch can be tolerated.

Therefore, the probes of the subject invention include mutations (both single and multiple), deletions, insertions f the described sequences, and combinations thereof, wherein said mutations, insertions and deletions permit formation of stable hybrids with the target polynucleotide of interest. Mutations, insertions, and deletions can be produced in a given polynucleotide sequence in many ways, and these methods are known to an ordinarily skilled artisan. Other methods may become known in the future.

The known methods include, but are not limited to:

(1) synthesizing chemically or otherwise an artificial sequence which is a mutation, insertion or deletion of the known sequence;

(2) using a probe of the present invention to obtain via hybridization a new sequence or a mutation, insertion or deletion of the probe sequence; and

(3) mutating, inserting or deleting a test sequence in vitro or in vivo.

It is important to note that the mutational, insertional, and deletional variants generated from a given probe may be more or less efficient than the original probe. Notwithstanding such differences in efficiency, these variants are within the scope of the present invention.

Thus, mutational, insertional, and deletional variants of the disclosed test sequences can be readily prepared by methods which are well known to those skilled in the art. These variants can be used in the same manner as the instant probes so long as the variants have substantial sequence homology with the probes. As used herein, substantial sequence homology refers to homology which is sufficient to enable the variant to function in the same capacity as the original probe. Preferably, this homology is greater than 50%; more preferably, this homology is greater than 75%; and most preferably, this homology is greater than 90%. The degree of homology needed for the variant to function in its intended capacity will depend upon the intended use of the sequence. It is well within the skill of a person trained in this art to make mutational, insertional, and deletional mutations which are designed to improve the function of the sequence or otherwise provide a methodological advantage.

Specific nucleotide probes useful, according to the subject invention, in the rapid identification of acaride-active genes can be prepared utilizing the sequence information provided herein.

The potential variations in the probes listed is due, in part, to the redundancy of the genetic code. Because of the redundancy of the genetic code, i.e., more than one coding nucleotide triplet (codon) can be used for most of the amino acids used to make proteins. Therefore different nucleotide sequences can code for a particular amino acid. Thus, the amino acid sequences of the *B.t.* toxins and peptides can be prepared by equivalent nucleotide sequences encoding the same amino acid sequence of the protein or peptide. Accordingly, the subject invention includes such equivalent nucleotide sequences. Also, inverse or complement sequences are an aspect of the subject invention and can be readily used by a person skilled in this art. In addition it has been shown that proteins f identified structure and function

may be constructed by changing the amino acid sequence if such changes do not alter the protein secondary structure (Kaiser, E.T. and Kezdy, F.J. [1984] Science 223:249-255). Thus, the subject invention includes mutants of the amino acid sequence depicted herein which do not alter the protein secondary structure, or if the structure is altered, the biological activity is substantially retained. Further, the invention also includes mutants of organisms hosting all or part of a toxin encoding a gene of the invention. Such microbial mutants can be made by techniques well known to persons skilled in the art. For example, UV irradiation can be used to prepare mutants of host organisms. Likewise, such mutants may include asporogenous host cells which also can be prepared by procedures well known in the art.

The B.t. isolates of the invention, and mutants thereof, can be cultured using standard known media and fermentation techniques. Upon completion of the fermentation cycle, the bacteria can be harvested by first separating the B.t. spores and crystals from the fermentation broth by means well known in the art. The recovered B.t. spores and crystals can be formulated into a wettable powder, a liquid concentrate, granules or other formulations by the addition of surfactants, dispersants, inert carriers and other components to facilitate handling and application for particular target pests. The formulation and application procedures are all well known in the art and are used with commercial strains. The novel B.t. isolates, and mutants thereof, can be used to control target pests.

The cultures of the subject invention were deposited in the Agricultural Research Service Patent Culture Collection (NRRL), Northern Regional Research Center, 1815 North University Street, Peoria, Illinois, 61604 USA.

Culture	Accession No.	Deposit Date
B.t. PS50C	NRRL B-18746	January 9, 1991
B.t. PS86A1	NRRL B-18400	August 16, 1988
B.t. PS69D1	NRRL B-18247	July 28, 1987
B.t. PS72L1	NRRL B-18780	March 7, 1991
B.t. PS75J1	NRRL B-18781	March 7, 1991
B.t. PS83E5	NRRL B-18782	March 7, 1991
B.t. PS45B1	NRRL B-18396	August 16, 1988
B.t. PS24J	NRRL B-18881	August 30, 1991
B.t. PS94R3	NRRL B-18882	August 30, 1991
B.t. PS17	NRRL B-18243	July 28, 1987
B.t. PS62B1	NRRL B-18398	August 16, 1988
B.t. PS74G1	NRRL B-18397	August 16, 1988
E. coli NM522(pMYC 2321)	NRRL B-18770	February 14, 1991
E. coli NM522(pMYC 2317)	NRRL B-18816	April 24, 1991
E. coli NM522(pMYC 1627)	NRRL B-18651	May 11, 1990
E. coli NM522(pMYC 1628)	NRRL B-18652	May 11, 1990
E. coli NM522(pMYC 1638)	NRRL B-18751	January 11, 1991

E. coli NM522(pMYC 1638)	NRRL B-18769	February 14, 1991

The subject cultures have been deposited under conditions that assure that access to the cultures will be available during the pendency of this patent application to one determined by the Commissioner of Patents and Trademarks to be entitled thereto under 37 CFR 1.14 and 35 U.S.C. 122. These deposits are available as required by foreign patent laws in countries wherein counterparts of the subject application, or its progeny, are filed. However, it should be understood that the availability of a deposit does not constitute a license to practice the subject invention in derogation of patent rights granted by governmental action.

Further, the subject culture deposits will be stored and made available to the public in accord with the provisions of the Budapest Treaty for the Deposit of Microorganisms, i.e., they will be stored with all the care necessary to keep them viable and uncontaminated for a period of at least five years after the most recent request for the furnishing of a sample of a deposit, and in any case, for a period of at least thirty (30) years after the date of deposit or for the enforceable life of any patent which may issue disclosing a culture. The depositor acknowledges the duty to replace a deposit should the depository be unable to furnish a sample when requested, due to the condition of a deposit. All restrictions on the availability to the public of the subject culture deposits will be irrevocably removed upon the granting of a patent disclosing them.

Upon applying an acaricidal-effective amount of a microbe, or toxin, as disclosed herein, in a suitable acaricidal formulation to the environment of the target pest, there is obtained effective control of these pests. An acaricidal-effective amount can vary from about 1 to about 12 l/ha, depending upon the nature and quantity of the pests to be controlled, the time of year, temperature, humidity, and other known factors which may affect a bioinsecticide. It is well within the skill of those trained in this art to determine the quantity of bioinsecticide to apply in order to obtain effective control of target pests.

The intracellular δ-endotoxin protein can be combined with other insecticidal proteins (including those obtained from sources other than Bacillus thuringiensis) to increase the spectrum of activity to give complete control of target pests.

The B.t. cells may be formulated in a variety of ways. They may be employed as wettable powders, granules or dusts, by mixing with various inert materials, such as inorganic minerals (phyllosilicates, carbonates, sulfates, phosphates, and the like) or botanical materials (powdered corncobs, rice hulls, walnut shells, and the like). The formulations may include spreader-sticker adjuvants, stabilizing agents, other pesticidal additives, or surfactants. Liquid formulations may be aqueous-based or non-aqueous and employed as foams, gels, suspensions, emulsifiable concentrates, or the like. The ingredients may include rheological agents, surfactants, emulsifiers, dispersants, r polymers.

The pesticidal concentration will vary widely depending upon the nature of the particular formulation, particularly whether it is a concentrate or to be used directly. The

pesticide will be present in at least 1% by weight and may be 100% by weight. The dry formulations will have from about 1-95% by weight of the pesticide while the liquid formulations will generally be from about 1-60% by weight of the solids in the liquid phase. The formulations will generally have from about 10^2 to about 10^4 cells/mg. These formulations will be administered at about 50 mg (liquid or dry) to 1 kg or more per hectare.

The formulations can be applied to the environment of the target pest(s), e.g., plants, livestock, fowl, soil or water, by spraying, dusting, sprinkling, or the like.

The toxin genes harbored by the novel isolates of the subject invention can be introduced into a wide variety of microbial hosts. Expression of the toxin gene results, directly or indirectly, in the intracellular production and maintenance of the pesticide. With suitable hosts, e.g., Pseudomonas, the microbes can be applied to the situs of mites where they will proliferate and be ingested by the mites. The result is a control of the mites. Alternatively, the microbe hosting the toxin gene can be treated under conditions that prolong the activity of the toxin produced in the cell. The treated cell then can be applied to the environment of the target pest. The resulting product retains the toxicity of the B.t. toxin.

Where the B.t. toxin gene is introduced via a suitable vector into a microbial host, and said host is applied to the environment in a living state, it is essential that certain host microbes be used. Microorganism hosts are selected which are known to occupy the "phytosphere" (phylloplane, phyllosphere, rhizosphere, and/or rhizoplane) of one or more crops of interest. These microorganisms are selected so as to be capable of successfully competing in the particular environment (crop and other insect habitats) with the wild-type microorganisms, provide for stable maintenance and expression of the gene expressing the polypeptide pesticide, and, desirably, provide for improved protection of the pesticide from environmental degradation and inactivation.

A large number of microorganisms are known to inhabit the phylloplane (the surface of the plant leaves) and/or the rhizosphere (the soil surrounding plant roots). These microorganisms include bacteria, algae, and fungi. Of particular interest are microorganisms, such as bacteria, e.g., genera Bacillus, Pseudomonas, Erwinia, Serratia, Klebsiella, Xanthomonas, Streptomyces, Rhizobium, Rhodopseudomonas, Methylophilius, Agrobacterium, Acetobacter, Lactobacillus, Arthrobacter, Azotobacter, Leuconostoc, Alcaligenes and Clostridium; fungi, particularly yeast, e.g., genera Saccharomyces, Cryptococcus, Kluyveromyces, Sporobolomyces, Rhodotorula, and Aureobasidium; microalgae, e.g., families Cyanophyceae, Prochlorophyceae, Rhodophyceae, Dinophyceae, Chrysophyceae, Prymnesiophyceae, Xanthophyceae, Raphidophyceae, Bacillariophyceae, Eustigmatophyceae, Cryptophyceae, Euglenophyceae, Prasinophyceae, and Chlorophyceae. Of particular interest are such phytosphere bacterial species as Pseudomonas syringae, Pseudomonas fluorescens, Serratia marcescens, Acetobacter xylinum, Agrobacterium tumefaciens, Rhodopseudomonas spheroides, Xanthomonas campestris, Rhizobium melioti, Alcaligenes entrophus, and Azotobacter vinlandii; and phytosphere yeast species such as Rhodotorula rubra, R. glutinis, R. marina, R.

aurantiaca, Cryptococcus albidus, C. diffluens, C. laurentii, Saccharomyces rosei, S. pretoriensis, S. cerevisiae, Sporobolomyces roseus, S. odorus, Kluyveromyces veronae, and Aureobasidium pollulans. Of particular interest are the pigmented microorganisms.

A wide variety of ways are available for introducing a B.t. gene expressing a toxin into the microorganism host under conditions which allow for stable maintenance and expression of the gene. One can provide for DNA constructs which include the transcriptional and translational regulatory signals for expression of the toxin gene, the toxin gene under their regulatory control and a DNA sequence homologous with a sequence in the host organism, whereby integration will occur, and/or a replication system which is functional in the host, whereby integration or stable maintenance will occur.

The transcriptional initiation signals will include a promoter and a transcriptional initiation start site. In some instances, it may be desirable to provide for regulative expression of the toxin, where expression of the toxin will only occur after release into the environment. This can be achieved with operators or a region binding to an activator or enhancers, which are capable of induction upon a change in the physical or chemical environment of the microorganisms. For example, a temperature sensitive regulatory region may be employed, where the organisms may be grown up in the laboratory without expression of a toxin, but upon release into the environment, expression would begin. Other techniques may employ a specific nutrient medium in the laboratory, which inhibits the expression of the toxin, where the nutrient medium in the environment would allow for expression of the toxin. For translational initiation, a ribosomal binding site and an initiation codon will be present.

Various manipulations may be employed for enhancing the expression of the messenger RNA, particularly by using an active promoter, as well as by employing sequences, which enhance the stability of the messenger RNA. The transcriptional and translational termination region will involve stop codon(s), a terminator region, and optionally, a polyadenylation signal. A hydrophobic "leader" sequence may be employed at the amino terminus of the translated polypeptide sequence in order to promote secretion of the protein across the inner membrane.

In the direction of transcription, namely in the 5' to 3' direction of the coding or sense sequence, the construct will involve the transcriptional regulatory region, if any, and the promoter, where the regulatory region may be either 5' or 3' of the promoter, the ribosomal binding site, the initiation codon, the structural gene having an open reading frame in phase with the initiation codon, the stop codon(s), the polyadenylation signal sequence, if any, and the terminator region. This sequence as a double strand may be used by itself for transformation of a microorganism host, but will usually be included with a DNA sequence involving a marker, where the second DNA sequence may be joined to the toxin expression construct during introduction of the DNA into the host.

By a marker is intended a structural gene which provides for selection of those hosts which have been modified or transformed. The marker will normally provide for

selective advantage, for example, providing for biocide resistance, e.g., resistance to antibiotics or heavy metals; complementation, so as to provide prototropy to an auxotrophic host, or the like. Preferably, complementation is employed, so that the modified host may not only be selected, but may also be competitive in the field. One or more markers may be employed in the development of the constructs, as well as for modifying the host. The organisms may be further modified by providing for a competitive advantage against other wild-type microorganisms in the field. For example, genes expressing metal chelating agents, e.g., siderophores, may be introduced into the host along with the structural gene expressing the toxin. In this manner, the enhanced expression of a siderophore may provide for a competitive advantage for the toxin-producing host, so that it may effectively compete with the wild-type microorganisms and stably occupy a niche in the environment.

Where no functional replication system is present, the construct will also include a sequence of at least 50 basepairs (bp), preferably at least about 100 bp, and usually not more than about 5000 bp of a sequence homologous with a sequence in the host. In this way, the probability of legitimate recombination is enhanced, so that the gene will be integrated into the host and stably maintained by the host. Desirably, the toxin gene will be in close proximity to the gene providing for complementation as well as the gene providing for the competitive advantage. Therefore, in the event that a toxin gene is lost, the resulting organism will be likely to also lose the complementing gene and/or the gene providing for the competitive advantage, so that it will be unable to compete in the environment with the gene retaining the intact construct.

A large number of transcriptional regulatory regions are available from a wide variety of microorganism hosts, such as bacteria, bacteriophage, cyanobacteria, algae, fungi, and the like. Various transcriptional regulatory regions include the regions associated with the trp gene, lac gene, gal gene, the lambda left and right promoters, the tac promoter, the naturally-occurring promoters associated with the toxin gene, where functional in the host. See for example, U.S. Patent Nos. 4,332,898, 4,342,832 and 4,356,270. The termination region may be the termination region normally associated with the transcriptional initiation region or a different transcriptional initiation region, so long as the two regions are compatible and functional in the host.

Where stable episomal maintenance or integration is desired, a plasmid will be employed which has a replication system which is functional in the host. The replication system may be derived from the chromosome, an episomal element normally present in the host or a different host, or a replication system from a virus which is stable in the host. A large number of plasmids are available, such as pBR322, pACYC184, RSF1010, pRO1614, and the like. See for example, Olson et al., (1982) J. Bacteriol. 150:6069, and Bagdasarian et al., (1981) Gene 16:237, and U.S. Patent Nos. 4,356,270, 4,362,817, and 4,371,625.

The B.t. gene can be introduced between the transcriptional and translational initiation region and the transcriptional and translational termination region, so as to be under

the regulatory control of the initiation region. This construct will be included in a plasmid, which will include at least ne replication system, but may include more than one, where one replication system is employed for cloning during the development of the plasmid and the second replication system is necessary for functioning in the ultimate host. In addition, one or more markers may be present, which have been described previously. Where integration is desired, the plasmid will desirably include a sequence homologous with the host genome.

The transformants can be isolated in accordance with conventional ways, usually employing a selection technique, which allows for selection of the desired organism as against unmodified organisms or transferring organisms, when present. The transformants then can be tested for pesticidal activity.

Suitable host cells, where the pesticide-containing cells will be treated to prolong the activity of the toxin in the cell when the then treated cell is applied to the environment of target pest(s), may include either prokaryotes or eukaryotes, normally being limited to those cells which do not produce substances toxic to higher organisms, such as mammals. However, organisms which produce substances toxic to higher organisms could be used, where the toxin is unstable or the level of application sufficiently low as to avoid any possibility of toxicity to a mammalian host. As hosts, of particular interest will be the prokaryotes and the lower eukaryotes, such as fungi, as disclosed previously.

Characteristics of particular interest in selecting a host cell for purposes of production include ease of introducing the B.t. gene into the host, availability of expression systems, efficiency of expression, stability of the pesticide in the host, and the presence of auxiliary genetic capabilities. Characteristics of interest for use as a pesticide microcapsule include protective qualities for the pesticide, such as thick cell walls, pigmentation, and intracellular packaging or formation of inclusion bodies; survival in aqueous environments; lack of mammalian toxicity; attractiveness to pests for ingestion; ease of killing and fixing without damage to the toxin; and the like. Other considerations include ease of formulation and handling, economics, storage stability, and the like.

The cell will usually be intact and be substantially in the proliferative form when treated, rather than in a spore form, although in some instances spores may be employed.

Treatment of the microbial cell, e.g., a microbe containing the B.t. toxin gene, can be by chemical or physical means, or by a combination of chemical and/or physical means, so long as the technique does not deleteriously affect the properties of the toxin, nor diminish the cellular capability in protecting the toxin. Examples of chemical reagents are halogenating agents, particularly halogens of atomic no. 17-80. More particularly, iodine can be used under mild conditions and for sufficient time to achieve the desired results. Other suitable techniques include treatment with aldehydes, such as formaldehyde and glutaraldehyde; anti-infectives, such as zephiran chloride and cetylpyridinium chloride; alcohols, such as isopropyl and ethanol; various histologic fixatives, such as Lugol iodine, Bouin's fixative, and Helly's fixative (See: Humason, Gretchen L., Animal Tissue Techniques, W.H. Freeman and

Company, 1967); or a combination of physical (heat) and chemical agents that preserve and prolong the activity of the toxin produced in the cell when the cell is administered to the host animal. Examples of physical means are short wavelength radiation such as gamma-radiation and X-radiation, freezing, UV irradiation, lyophilization, and the like.

The cells generally will have enhanced structural stability which will enhance resistance to environmental conditions. Where the pesticide is in a proform, the method of inactivation should be selected so as not to inhibit processing of the proform to the mature form of the pesticide by the target pest pathogen. For example, formaldehyde will crosslink proteins and could inhibit processing of the proform of a polypeptide pesticide. The method of inactivation or killing retains at least a substantial portion of the bio-availability or bioactivity of the toxin.

The cellular host containing the B.t. insecticidal gene may be grown in any convenient nutrient medium, where the DNA construct provides a selective advantage, providing for a selective medium so that substantially all or all of the cells retain the B.t. gene. These cells may then be harvested in accordance with conventional ways. Alternatively, the cells can be treated prior to harvesting.

The B.t. cells of the invention can be cultured using standard art media and fermentation techniques. Upon completion of the fermentation cycle the bacteria can be harvested by first separating the B.t. spores and crystals from the fermentation broth by means well known in the art. The recovered B.t. spores and crystals can be formulated into a wettable powder, liquid concentrate, granules or other formulations by the addition of surfactants, dispersants, inert carriers, and other components to facilitate handling and application for particular target pests. These formulations and application procedures are all well known in the art.

Formulated bait granules containing an attractant and spores and crystals of the B.t. isolates, or recombinant microbes comprising the gene(s) obtainable from the B.t. isolates disclosed herein, can be applied to the soil or in the vicinity of stored products. Formulated product can also be applied as a seed-coating or root treatment or total plant treatment at later stages of the crop cycle.

Mutants of the novel isolates of the invention can be made by procedures well known in the art. For example, an asporogenous mutant can be obtained through ethylmethane sulfonate (EMS) mutagenesis of a novel isolate. The mutants can be made using ultraviolet light and nitrosoguanidine by procedures well known in the art.

A smaller percentage of the asporogenous mutants will remain intact and not lyse for extended fermentation periods; these strains are designated lysis minus (-). Lysis minus strains can be identified by screening asporogenous mutants in shake flask media and selecting those mutants that are still intact and contain toxin crystals at the end of the fermentation. Lysis minus strains are suitable for a cell fixation process that will yield a protected, encapsulated toxin protein.

To prepare a phage resistant variant of said asporogenous mutant, an aliquot of the phage lysate is spread onto nutrient agar and allowed to dry. An aliquot of the phage sensitive bacterial strain is then plated directly over the dried lysate and allowed to dry. The plates are incubated at 30°C. The plates are incubated for 2 days and, at that time, numerous colonies could be seen growing on the agar. Some of these colonies are picked and subcultured onto nutrient agar plates. These apparent resistant cultures are tested for resistance by cross streaking with the phage lysate. A line of the phage lysate is streaked on the plate and allowed to dry. The presumptive resistant cultures are then streaked across the phage line. Resistant bacterial cultures show no lysis anywhere in the streak across the phage line after overnight incubation at 30°C. The resistance to phage is then reconfirmed by plating a lawn of the resistant culture onto a nutrient agar plate. The sensitive strain is also plated in the same manner to serve as the positive control. After drying, a drop of the phage lysate is plated in the center of the plate and allowed to dry. Resistant cultures showed no lysis in the area where the phage lysate has been placed after incubation at 30°C for 24 hours.

Following are examples which illustrate procedures, including the best mode, for practicing the invention. These examples should not be construed as limiting. All percentages are by weight and all solvent mixture proportions are by volume unless otherwise noted.

Example 1 – Culturing of the *B.t.* Isolates

A subculture of the B.t. isolates, or mutants thereof, can be used to inoculate the following medium, a peptone, glucose, salts medium.

Bacto Peptone	7.5 g/l
Glucose	1.0 g/l
KH_2PO_4	3.4 g/l
K_2HPO_4	4.35 g/l
Salt Solution	5.0 ml/l
$CaCl_2$ Solution	5.0 ml/l
pH 7.2	

Salts Solution (100 ml)

$MgSO_4.7H_2O$	2.46 g
$MnSO_4.H_2O$	0.04 g
$ZnSO_4.7H_2O$	0.28 g
$FeSO_4.7H_2O$	0.40 g

$CaCl_2$ Solution (100 ml)

$CaCl_2.2H_2O$	3.66 g

The salts solution and $CaCl_2$ solution are filter-sterilized and added to the autoclaved and cooked broth at the time of inoculation. Flasks are incubated at 30°C on a rotary shaker at 200 rpm for 64 hr.

The above procedure can be readily scaled up to large fermentors by procedures well known in the art.

The B.t. spores and/or crystals, obtained in the above fermentation, can be isolated by procedures well known in the art. A frequently-used procedure is to subject the harvested fermentation broth to separation techniques, e.g., centrifugation.

Example 2 – Purification of Protein and Amino Acid Sequencing

The B.t. isolates PS17, PS52A1 and PS69D1 were cultured as described in Example 1. The parasporal inclusion bodies were partially purified by sodium bromide (28-38%) isopycnic gradient centrifugation (Pfannenstiel, M.A., E.J. Ross, V.C. Kramer, and K.W. Nickerson [1984] FEMS Microbiol. Lett. 21:39). The proteins were bound to PVDF membranes (Millipore, Bedford, MA) by western blotting techniques (Towbin, H., T. Staehlelin, and K. Gordon [1979] Proc. Natl. Acad. Sci. USA 76:4350) and the N-terminal amino acid sequences were determined by the standard Edman reaction with an automated gas-phase sequenator (Hunkapiller, M.W., R.M. Hewick, W.L. Dreyer, and L.E. Hood [1983] Meth. Enzymol. 91:399). The sequences obtained were:

PS17a: A I L N E L Y P S V P Y N V (SEQ ID NO. 12)

PS17b: A I L N E L Y P S V P Y N V (SEQ ID NO. 13)

PS52A1: M I I D S K T T L P R H S L I N T (SEQ ID NO. 14)

PS69D1: M I L G N G K T L P K H I R L A H I F A T Q N S (SEQ ID NO. 15)

Example 3 – Cloning of Novel Toxin Genes and Transformation into Escherichia coli

Total cellular DNA was prepared by growing the cells *B.t.* PS17 to a low optical density (OD_{600} = 1.0) and recovering the cells by centrifugation. The cells were protoplasted in TES buffer (30 mM Tris-Cl, 10 mM EDTA, 50 mM NaCl, pH = 8.0) containing 20 % sucrose and 50 mg/ml lysozyme. The protoplasts were lysed by addition of SDS to a final concentration of 4%. The cellular material was precipitated overnight at 4°C in 100 mM (final concentration) neutral potassium chloride. The supernate was extracted twice with phenol/chloroform (1:1). The DNA was precipitated with ethanol and purified by isopycnic banding on a cesium chloride-ethidium bromide gradient.

Total cellular DNA from PS17 was digested with EcoRI and separated by electrophoresis on a 0.8% (w/v) Agarose-TAE (50 mM Tris-HCl, 20 mM NaOAc, 2.5 mM EDTA, pH=8.0) buffered gel. A Southern blot of the gel was hybridized with a [^{32}P] - radiolabeled oligonucleotide probe derived from the N-terminal amino acid sequence of purified 130 kDa protein from PS17. The sequence of the oligonucleotide synthesized is (GCAATTTTAAATGAATTATATCC) (SEQ ID NO. 16). Results showed that the

hybridizing EcoRI fragments of PS17 are 5.0 kb, 4.5 kb, 2.7 kb and 1.8 kb in size, presumptively identifying at least four new acaride-active toxin genes, PS17d, PS17b, PS17a and PS17e, respectively.

A library was constructed from PS17 total cellular DNA partially digested with Sau3A and size fractionated by electrophoresis. The 9 to 23 kb region of the gel was excised and the DNA was electroeluted and then concentrated using an Elutip™ ion exchange column (Schleicher and Schuel, Keene NH). The isolated Sau3A fragments were ligated into LambdaGEM-11™ (PROMEGA). The packaged phage were plated on KW251 E. coli cells (PROMEGA) at a high titer and screened using the above radiolabeled synthetic oligonucleotide as a nucleic acid hybridization probe. Hybridizing plaques were purified and rescreened at a lower plaque density. Single isolated purified plaques that hybridized with the probe were used to infect KW251 E. coli cells in liquid culture for preparation of phage for DNA isolation. DNA was isolated by standard procedures.

Recovered recombinant phage DNA was digested with EcoRI and separated by electrophoresis on a 0.8% agarose-TAE gel. The gel was Southern blotted and hybridized with the oligonucleotide probe to characterize the toxin genes isolated from the lambda library. Two patterns were present, clones containing the 4.5 kb (PS17b) or the 2.7 kb (PS17a) EcoRI fragments. Preparative amounts of phage DNA were digested with SalI (to release the inserted DNA from lambda arms) and separated by electrophoresis on a 0.6% agarose-TAE gel. The large fragments, electroeluted and concentrated as described above, were ligated to SalI-digested and dephosphorylated pBClac, an E. coli/B.t. shuttle vector comprised of replication origins from pBC16 and pUC19 . The ligation mix was introduced by transformation into NM522 competent E. coli cells and plated on LB agar containing ampicillin, isopropyl-(Beta)-D-thiogalactoside (IPTG) and 5-Bromo-4-Chloro-3-indolyl-(Beta)-D-galactoside (XGAL). White colonies, with putative insertions in the (Beta)-galactosidase gene of pBClac, were subjected to standard rapid plasmid purification procedures to isolate the desired plasmids. The selected plasmid containing the 2.7 kb EcoRI fragment was named pMYC1627 and the plasmid containing the 4.5 kb EcoRI fragment was called pMYC1628.

The toxin genes were sequenced by the standard Sanger dideoxy chain termination method using the synthetic oligonucleotide probe, disclosed above, and by "walking" with primers made to the sequence of the new toxin genes.

The PS17 toxin genes were subcloned into the shuttle vector pHT3101 (Lereclus, D. et al. [1989] FEMS Microbiol. Lett. 60:211-218) using standard methods for expression in B.t. Briefly, SalI fragments containing the 17a and 17b toxin genes were isolated from pMYC1629 and pMYC1627, respectively, by preparative agarose gel electrophoresis, electroelution, and concentrated, as described above. These concentrated fragments were ligated into SalI-cleaved and dephosphorylated pHT3101. The ligation mixtures were used separately to transform frozen, competent E. coli NM522. Plasmids from each respective recombinant E. coli strain were prepared by alkaline lysis and analyzed by agarose gel

electrophoresis. The resulting subclones, pMYC2311 and pMYC2309, harbored the 17a and 17b toxin genes, respectively. These plasmids were transformed into the acrystalliferous B.t. strain, HD-1 cryB (Aronson, A., Purdue University, West Lafayette, IN), by standard electroporation techniques (Instruction Manual, Biorad, Richmond, CA).

Recombinant B.t. strains HD-1 cryB [pMYC2311] and [pMYC2309] were grown to sporulation and the proteins purified by NaBr gradient centrifugation as described above for the wild-type B.t. proteins.

Example 4 – Molecular Cloning of Gene Encoding a Novel Toxin From Bacillus thuringiensis strain PS52A1

Total cellular DNA was prepared from *Bacillus thuringiensis* PS52A1 (B.t. PS52A1) as disclosed in Example 3.

RFLP analyses were performed by standard hybridization of Southern blots of PS52A1 DNA with a ^{32}P-labeled oligonucleotide probe designed from the N-terminal amino acid sequence disclosed in Example 2. The sequence of this probe is:

5' ATG ATT ATT GAT TCT AAA ACA ACA TTA CCA AGA CAT TCA/T TTA ATA/T AAT ACA/T ATA/T AA 3' (SEQ ID NO. 17)

This probe was designated 52A1-C. Hybridizing bands included an approximately 3.6 kbp *Hin*dIII fragment and an approximately 8.6 kbp *Eco*RV fragment. A gene library was constructed from PS52A1 DNA partially digested with *Sau*3A. Partial restriction digests were fractionated by agarose gel electrophoresis. DNA fragments 6.6 to 23 kbp in size were excised from the gel, electroeluted from the gel slice, and recovered by ethanol precipitation after purification on an Elutip-D ion exchange column. The *Sau*3A inserts were ligated into *Bam*HI-digested LambdaGem-11 (Promega). Recombinant phage were packaged and plated on *E. coli* KW251 cells (Promega). Plaques were screened by hybridization with the radiolabeled 52A1-C oligonucleotide probe disclosed above. Hybridizing phage were plaque-purified and used to infect liquid cultures of *E. coli* KW251 cells for isolation of phage DNA by standard procedures (Maniatis et al.). For subcloning, preparative amounts of DNA were digested with *Eco*RI and *Sal*I, and electrophoresed on an agarose gel. The approximately 3.1 kbp band containing the toxin gene was excised from the gel, electroeluted from the gel slice, and purified by ion exchange chromatography as above. The purified DNA insert was ligated into *Eco*RI + *Sal*I-digested pHTBlueII (an *E. coli/B. thuringiensis* shuttle vector comprised of pBluescript S/K [Stratagene] and the replication origin from a resident *B.t.* plasmid [D. Lereclus et al. 1989. FEMS Microbiology Letters 60:211-218]). The ligation mix was used to transform frozen, competent *E. coli* NM522 cells (ATCC 47000). Transformants were plated on LB agar containing ampicillin, isopropyl-(Beta)-D-thiogalactoside (IPTG), and 5-Bromo-4-Chloro-3-indolyl-(Beta)-D-galactoside (XGAL). Plasmids were purified from putative recombinants by alkaline lysis (Maniatis et al.) and analyzed by electrophoresis of *Eco*RI and

*Sal*I digests on agarose gels. The desired plasmid construct, pMYC2321 contains a toxin gene that is novel compared to the maps of other toxin genes encoding acaricidal proteins.

Plasmid pMYC2321 was introduced into an acrystalliferous (Cry⁻) *B.t.* host by electroporation. Expression of an approximately 55-60 kDa crystal protein was verified by SDS-PAGE analysis.

Example 5 – Molecular Cloning of Gene Encoding a Novel Toxin From *Bacillus Thuringiensis* strain PS69D1

Total cellular DNA was prepared from PS69D1 (*B.t.* PS69D1) as disclosed in Example 3. RFLP analyses were performed by standard hybridization of Southern blots of PS69D1 DNA with a 32P-labeled oligonucleotide probe designated as 69D1-D. The sequence of the 69D1-D probe was:

5' AAA CAT ATT AGA TTA GCA CAT ATT TTT GCA ACA CAA AA 3' (SEQ ID NO. 18)

Hybridizing bands included an approximately 2.0 kbp *Hin*dIII fragment.

A gene library was constructed from PS69D1 DNA partially digested with *Sau*3A. Partial restriction digests were fractionated by agarose gel electrophoresis. DNA fragments 6.6 to 23 kbp in size were excised from the gel, electroeluted from the gel slice, and recovered by ethanol precipitation after purification on an Elutip-D ion exchange column. The *Sau*3A inserts were ligated into *Bam*HI-digested LambdaGem-11 (Promega, Madison, WI). Recombinant phage were packaged and plated on *E. coli* KW251 cells (Promega, Madison, WI). Plaques were screened by hybridization with the radiolabeled 69D1-D oligonucleotide probe. Hybridizing phage were plaque-purified and used to infect liquid cultures of *E. coli* KW251 cells for isolation of phage DNA by standard procedures (Maniatis et al. [1982] *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor, NY). For subcloning, preparative amounts of DNA were digested with *Hin*dIII and electrophoresed on an agarose gel. The approximately, 2.0 kbp band containing the toxin gene was excised from the gel, electroeluted from the gel slice, and purified by ion exchange chromatography as above. The purified DNA insert was ligated into *Hin*dIII-digested pHTBlueII (and *E. coli*/*B.t.* shuttle vector comprised of pBluescript S/K (Stratagene, San Diego, CA) and the replication origin from a resident *B.t.* plasmid (D. Lereclus et al [1989] FEMS Microbiol. Lett. 60:211-218). The ligation mix was used to transform frozen, competent *E. coli* NM522 cells (ATCC 47000). Transformants were plated on LB agar containing 5-bromo-4-chloro-3-indolyl-(Beta)-D-galactoside (XGAL). Plasmids were purified from putative recombinants by alkaline lysis (Maniatis et al., *supra*) and analyzed by electrophoresis of *Hin*dIII digests on agarose gels. The desired plasmid construct, pMYC2317, contains a toxin gene that is novel compared to the maps of other toxin genes encoding insecticidal proteins.

Example 6 – Activity of B.t. Isolates Against Mites

B. thuringiensis isolates of the invention were tested as spray-dried powders of fermentation broths which were concentrated by centrifugation. Pellets, which consist of water and biomass (spores, crystalline delta-endotoxins, cellular debris and growth media) were mixed with a standard carrier, preservative and surfactant. Powders, which consisted of 25% biomass, were made using a Yamato spray drier. (Sold by Yamato Scientific Co., Ltd. Tokoyo, Japan)

All broths were tested for the presence of beta-exotoxin by a larval house fly bioassay (Campbell, D.P., Dieball, D.E. and Brackett, J.M., 1987, Rapid HPLC assay for the ß-exotoxin of Bacillus thuringiensis. J. Agric. Food Chem. 35:156-158). Only isolates which tested free of ß-exotoxin were used in the assays against mites.

B. thuringiensis isolates were tested using an artificial feeding assay. Spray-dried powders were prepared for testing by mixing 25mg of powder in 5 ml of a 10% sucrose solution. This mixture was then sonicated for 8 min to produce a suspension.

Two ml of suspension was placed in a reservoir consisting of a metal ring with a Parafilm™M film bottom. A petri dish containing approximately 30 female Two-spotted spider mites (Tetranychus urticae) was placed on the underside of the film. Mites were allowed to feed on the sucrose solution for 24 hrs and then transfered to 2 cm French bean leaf discs (20 mites per disc). Mortality was determined after 7 days (Table 2). Each assay was done in triplicate.

TABLE 2. Toxicity of Bacillus thuringiensis isolates to the two spotted spider mite, Tetranychus urticae. Mortality was determined after 7 days of treatment.

Isolate	Percent Mortality
B.t. PS50C	63
B.t. PS86A1	85
B.t. PS69D1	77
B.t. PS72L1	85
B.t. PS75J1	85
B.t. PS83E5	70
B.t. PS45B1	82
B.t. PS24J	90
B.t. PS94R3	97
B.t. PS17	>90
B.t. PS62B1	>90
B.t. PS74G1	>90
Control	10

Example 7 – Cloning of Novel Acaride-Active Genes Using Generic Oligonucleotide Primers

The acaricidal gene of a new acaricidal *B.t.* isolate can be obtained from DNA of the strain by performing the standard polymerase chain reaction using the oligonucleotides of SEQ ID NO. 21 or SEQ ID NO. 20 as reverse primers and SEQ ID NO. 10, SEQ ID NO. 11, SEQ ID NO. 16, Probe B of SEQ ID NO. 5 (AAT GAA GTA/T TAT CCA/T GTA/T AAT), or SEQ ID NO. 19 as forward primers. The expected PCR fragments would be approximately 330 to 600 bp (with either reverse primer and SEQ ID NO. 10), 1000 to 1400 bp (with either reverse primer and SEQ ID NO. 11), and 1800 to 2100 bp (with either reverse primer and any of the three N-terminal primers, SEQ ID NO. 5 (Probe B), SEQ ID NO. 16, and SEQ ID NO. 19). Alternatively, a complement from the primer family described by SEQ ID NO. 10 can be used as reverse primer with SEQ ID NO. 11, SEQ ID NO. 16, SEQ ID NO. 5 (Probe B), or SEQ ID NO. 19 as forward primers. The expected PCR fragments would be approximately 650 to 1000 bp with SEQ ID NO. 11, and 1400 to 1800 bp (for the three N-terminal primers, SEQ ID NO. 5 (Probe B), SEQ ID NO. 16, and SEQ ID NO. 19). Amplified DNA fragments of the indicated sizes can be radiolabeled and used as probes to clone the entire gene.

Example 8 – Further Cloning of Novel Acaride-Active Genes Using Generic Oligonucleotide Primers

A gene coding for a acaricidal toxin of an acaricidal *B.t.* isolate can also be obtained from DNA of the strain by performing the standard polymerase chain reaction using oligonucleotides derived from the PS52A1 and PS69D1 gene sequences as follows:

1. Forward primer "TGATTTT(T or A)(C or A)TCAATTATAT(A or G)A(G or T)GTTTAT" (SEQ ID NO. 22) can be used with primers complementary to probe "AAGAGTTA(C or T)TA(A or G)A(G or A)AAAGTA" (SEQ ID NO. 23), probe "TTAGGACCATT(A or G)(C or T)T(T or A)GGATTTGTTGT(A or T)TATGAAAT" (SEQ ID NO. 24), and probe "GA(C or T)AGAGATGT(A or T)AAAAT(C or T)(T or A)TAGGAATG" (SEQ ID NO. 25) to produce amplified fragments of approximately 440, 540, and 650 bp, respectively.

2. Forward primer "TT(A or C)TTAAA(A or T)C(A or T)GCTAATGATATT" (SEQ ID NO. 26) can be used with primers complementary to SEQ ID NO. 23, SEQ ID NO. 24, and SEQ ID NO. 25 to produce amplified fragments of approximately 360, 460, and 570 bp, respectively.

3. Forward primer SEQ ID NO. 23 can be used with primers complementary to SEQ ID NO. 24 and SEQ ID NO. 25 to produce amplified fragments of approximately 100 and 215 bp, respectively.

Amplified DNA fragments of the indicated sizes can be radiolabeled and used as probes to clone the entire gene.

Example 9 – Insertion of Toxin Genes Into Plants

One aspect of the subject invention is the transformation f plants with genes coding for a acaricidal toxin. The transformed plants are resistant to attack by acarides.

Genes coding for acaricidal toxins, as disclosed herein, can be inserted into plant cells using a variety of techniques which are well known in the art. For example, a large number of cloning vectors comprising a replication system in E. coli and a marker that permits selection of the transformed cells are available for preparation for the insertion of foreign genes into higher plants. The vectors comprise, for example, pBR322, pUC series, M13mp series, pACYC184, etc. Accordingly, the sequence coding for the B.t. toxin can be inserted into the vector at a suitable restriction site. The resulting plasmid is used for transformation into E. coli. The E. coli cells are cultivated in a suitable nutrient medium, then harvested and lysed. The plasmid is recovered. Sequence analysis, restriction analysis, electrophoresis, and other biochemical-molecular biological methods are generally carried out as methods of analysis. After each manipulation, the DNA sequence used can be cleaved and joined to the next DNA sequence. Each plasmid sequence can be cloned in the same or other plasmids. Depending on the method of inserting desired genes into the plant, other DNA sequences may be necessary. If, for example, the Ti or Ri plasmid is used for the transformation of the plant cell, then at least the right border, but often the right and the left border of the Ti or Ri plasmid T-DNA, has to be joined as the flanking region of the genes to be inserted.

The use of T-DNA for the transformation of plant cells has been intensively researched and sufficiently described in EP 120 516; Hoekema (1985) In: The Binary Plant Vector System, Offset-durkkerij Kanters B.V., Alblasserdam, Chapter 5; Fraley et al., Crit. Rev. Plant Sci. 4:1-46; and An et al. (1985) EMBO J. 4:277-287.

Once the inserted DNA has been integrated in the genome, it is relatively stable there and, as a rule, does not come out again. It normally contains a selection marker that confers on the transformed plant cells resistance to a biocide or an antibiotic, such as kanamycin, G 418, bleomycin, hygromycin, or chloramphenicol, inter alia. The individually employed marker should accordingly permit the selection of transformed cells rather than cells that do not contain the inserted DNA.

A large number of techniques are available for inserting DNA into a plant host cell. Those techniques include transformation with T-DNA using Agrobacterium tumefaciens or Agrobacterium rhizogenes as transformation agent, fusion, injection, or electroporation as well as other possible methods. If agrobacteria are used for the transformation, the DNA to be inserted has to be cloned into special plasmids, namely either into an intermediate vector or into a binary vector. The intermediate vectors can be integrated into the Ti or Ri plasmid by homologous recombination owing to sequences that are homologous to sequences in the T-DNA. The Ti or Ri plasmid also comprises the vir region necessary for the transfer of the

T-DNA. Intermediate vectors cannot replicate themselves in agrobacteria. The intermediate vector can be transferred into Agrobacterium tumefaciens by means of a helper plasmid (conjugation). Binary vectors can replicate themselves both in E. coli and in agrobacteria. They comprise a selection marker gene and a linker or polylinker which are framed by the right and left T-DNA border regions. They can be transformed directly into agrobacteria (Holsters et al. [1978] Mol. Gen. Genet. 163:181-187). The agrobacterium used as host cell is to comprise a plasmid carrying a vir region. The vir region is necessary for the transfer of the T-DNA into the plant cell. Additional T-DNA may be contained. The bacterium so transformed is used for the transformation of plant cells. Plant explants can advantageously be cultivated with Agrobacterium tumefaciens or Agrobacterium rhizogenes for the transfer of the DNA into the plant cell. Whole plants can then be regenerated from the infected plant material (for example, pieces of leaf, segments of stalk, roots, but also protoplasts or suspension-cultivated cells) in a suitable medium, which may contain antibiotics or biocides for selection. The plants so obtained can then be tested for the presence of the inserted DNA. No special demands are made of the plasmids in the case of injection and electroporation. It is possible to use ordinary plasmids, such as, for example, pUC derivatives.

The transformed cells grow inside the plants in the usual manner. They can form germ cells and transmit the transformed trait(s) to progeny plants. Such plants can be grown in the normal manner and crossed with plants that have the same transformed hereditary factors or other hereditary factors. The resulting hybrid individuals have the corresponding phenotypic properties.

Example 10 – Cloning of *Bacillus thuringiensis* Genes Into Baculoviruses

The genes coding for the insecticidal toxins, as disclosed herein, can be cloned into baculoviruses such as Autographa californica nuclear polyhedrosis virus (AcNPV). Plasmids can be constructed that contain the AcNPV genome cloned into a commercial cloning vector such as pUC8. The AcNPV genome is modified so that the coding region of the polyhedrin gene is removed and a unique cloning site for a passenger gene is placed directly behind the polyhedrin promoter. Examples of such vectors are pGP-B6874, described by Pennock et al. (Pennock, G.D., Shoemaker, C. and Miller, L.K. [1984] Mol. Cell. Biol. 4:399-406), and pAC380, described by Smith et al. (Smith, G.E., Summers, M.D. and Fraser, M.J. [1983] Mol Cell. Biol. 3:2156-2165). The genes coding for the protein toxins of the invention can be modified with BamHI linkers at appropriate regions both upstream and downstream from the coding region and inserted into the passenger site of one of the AcNPV vectors.

It should be understood that the examples and embodiments described herein are for illustrative purposes only and that various modifications or changes in light thereof will be suggested to persons skilled in the art and are to be included within the spirit and purview of this application and the scope of the appended claims.

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Payne, Jewel M.
Cannon, Raymond J.C.
Bagley, Angela L.

(ii) TITLE OF INVENTION: Novel *Bacillus thuringiensis* Isolates for Controlling Acarides

(iii) NUMBER OF SEQUENCES: 30

(iv) CORRESPONDENCE ADDRESS:
(A) ADDRESSEE: David R. Saliwanchik
(B) STREET: 2421 N.W. 41st Street, Suite A-1
(C) CITY: Gainesville
(D) STATE: FL
(E) COUNTRY: USA
(F) ZIP: 32606

(v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.25

(vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER: US
(B) FILING DATE:
(C) CLASSIFICATION:

(viii) ATTORNEY/AGENT INFORMATION:
(A) NAME: Saliwanchik, David R.
(B) REGISTRATION NUMBER: 31,794
(C) REFERENCE/DOCKET NUMBER: M/S 104

(ix) TELECOMMUNICATION INFORMATION:
(A) TELEPHONE: 904-375-8100
(B) TELEFAX: 904-372-5800

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 4155 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: Bacillus thuringiensis
(B) STRAIN: PS17
(C) INDIVIDUAL ISOLATE: PS17a

(vii) IMMEDIATE SOURCE:
(B) CLONE: E. coli NM522(pMYC 1627) NRRL B-18651

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

ATGGCAATTT TAAATGAATT ATATCCATCT GTACCTTATA ATGTATTGGC GTATACGCCA 60
CCCTCTTTTT TACCTGATGC GGGTACACAA GCTACACCTG CTGACTTAAC AGCTTATGAA 120
CAATTGTTGA AAAATTTAGA AAAAGGGATA AATGCTGGAA CTTATTCGAA AGCAATAGCT 180
GATGTACTTA AAGGTATTTT TATAGATGAT ACAATAAATT ATCAAACATA TGTAAATATT 240
GGTTTAAGTT TAATTACATT AGCTGTACCG GAAAATTGGTA TTTTTACACC TTTCATCGGT 300
TTGTTTTTTG CTGCATTGAA TAAACATGAT GCTCCACCTC CTCCTAATGC AAAAGATATA 360
TTTGAGGCTA TGAAACCAGC GATTCAAGAG ATGATTGATA GAACTTTAAC TGCGGATGAG 420
CAAACATTTT TAAATGGGGA AATAAGTGGT TTACAAAATT TAGCAGCAAG ATACCAGTCT 480
ACAATGGATG ATATTCAAAG CCATGGAGGA TTTAATAAGG TAGATTCTGG ATTAATTAAA 540
AAGTTTACAG ATGAGGTACT ATCTTTAAAT AGTTTTTATA CAGATCGTTT ACCTGTATTT 600

ATTACAGATA ATACAGCGGA TCGAACTTTG TTAGGTCTTC CTTATTATGC TATACTTGCG 660
AGCATGCATC TTATGTTATT AAGAGATATC ATTACTAAGG GTCCGACATG GGATTCTAAA 720
ATTAATTTCA CACCAGATGC AATTGATTCC TTTAAAACCG ATATTAAAAA TAATATAAAG 780
CTTTACTCTA AAACTATTTA TGACGTATTT CAGAAGGGAC TTGCTTCATA CGGAACGCCT 840
TCTGATTTAG AGTCCTTTGC AAAAAAACAA AAATATATTG AAATTATGAC AACACATTGT 900
TTAGATTTTG CAAGATTGTT TCCTACTTTT GATCCAGATC TTTATCCAAC AGGATCAGGT 960
GATATAAGTT TACAAAAAAC ACGTAGAATT CTTTCTCCTT TTATCCCTAT ACGTACTGCA 1020
GATGGGTTAA CATTAAATAA TACTTCAATT GATACTTCAA ATTGGCCTAA TTATGAAAAT 1080
GGGAATGGCG CGTTTCCAAA CCCAAAAGAA AGAATATTAA AACAATTCAA ACTGTATCCT 1140
AGTTGGAGAG CGGGACAGTA CGGTGGGCTT TTACAACCTT ATTTATGGGC AATAGAAGTC 1200
CAAGATTCTG TAGAGACTCG TTTGTATGGG CAGCTTCCAG CTGTAGATCC ACAGGCAGGG 1260
CCTAATTATG TTTCCATAGA TTCTTCTAAT CCAATCATAC AAATAAATAT GGATACTTGG 1320
AAAACACCAC CACAAGGTGC GAGTGGGTGG AATACAAATT TAATGAGAGG AAGTGTAAGC 1380
GGGTTAAGTT TTTTACAACG AGATGGTACG AGACTTAGTG CTGGTATGGG TGGTGGTTTT 1440
GCTGATACAA TATATAGTCT CCCTGCAACT CATTATCTTT CTTATCTCTA TGGAACTCCT 1500
TATCAAACTT CTGATAACTA TTCTGGTCAC GTTGGTGCAT TGGTAGGTGT GAGTACGCCT 1560
CAAGAGGCTA CTCTTCCTAA TATTATAGGT CAACCAGATG AACAGGGAAA TGTATCTACA 1620
ATGGGATTTC CGTTTGAAAA AGCTTCTTAT GGAGGTACAG TTGTTAAAGA ATGGTTAAAT 1680
GGTGCGAATG CGATGAAGCT TTCTCCTGGG CAATCTATAG GTATTCCTAT TACAAATGTA 1740
ACAAGTGGAG AATATCAAAT TCGTTGTCGT TATGCAAGTA ATGATAATAC TAACGTTTTC 1800
TTTAATGTAG ATACTGGTGG AGCAAATCCA ATTTTCCAAC AGATAAACTT TGCATCTACT 1860
GTAGATAATA ATACGGGAGT ACAAGGAGCA AATGGTGTCT ATGTAGTCAA ATCTATTGCT 1920
ACAACTGATA ATTCTTTTAC AGAAATTCCT GCGAAGACGA TTAATGTTCA TTTAACCAAC 1980
CAAGGTTCTT CTGATGTCTT TTTAGACCGT ATTGAATTTA TACCTTTTTC TCTACCTCTT 2040
ATATATCATG GAAGTTATAA TACTTCATCA GGTGCAGATG ATGTTTTATG GTCTTCTTCA 2100
AATATGAATT ACTACGATAT AATAGTAAAT GGTCAGGCCA ATAGTAGTAG TATCGCTAGT 2160
TCTATGCATT TGCTTAATAA AGGAAAAGTG ATAAAAACAA TTGATATTCC AGGGCATTCG 2220
GAAACCTTCT TTGCTACGTT CCCAGTTCCA GAAGGATTTA ATGAAGTTAG AATTCTTGCT 2280
GGCCTTCCAG AAGTTAGTGG AAATATTACC GTACAATCTA ATAATCCGCC TCAACCTAGT 2340
AATAATGGTG GTGGTGATGG TGGTGGTAAT GGTGGTGGTG ATGGTGGTCA ATACAATTTT 2400
TCTTTAAGCG GATCTGATCA TACGACTATT TATCATGGAA AACTTGAAAC TGGGATTCAT 2460
GTACAAGGTA ATTATACCTA TACAGGTACT CCCGTATTAA TACTGAATGC TTACAGAAAT 2520
AATACTGTAG TATCAAGCAT TCCAGTATAT TCTCCTTTTG ATATAACTAT ACAGACAGAA 2580
GCTGATAGCC TTGAGCTTGA ACTACAACCT AGATATGGTT TTGCCACAGT GAATGGTACT 2640
GCAACAGTAA AAAGTCCTAA TGTAAATTAC GATAGATCAT TTAAACTCCC AATAGACTTA 2700
CAAAATATCA CAACACAAGT AAATGCATTA TTCGCATCTG GAACACAAAA TATGCTTGCT 2760
CATAATGTAA GTGATCATGA TATTGAAGAA GTTGTATTAA AAGTGGATGC CTTATCAGAT 2820
GAAGTATTTG GAGATGAGAA GAAGGCTTTA CGTAAATTGG TGAATCAAGC AAAACGTTTG 2880
AGTAGAGCAA GAAATCTTCT GATAGGTGGG AGTTTTGAAA ATTGGGATGC ATGGTATAAA 2940
GGAAGAAATG TAGTAACTGT ATCTGATCAT GAACTATTTA AGAGTGATCA TGTATTATTA 3000
CCACCACCAG GATTGTCTCC ATCTTATATT TTCCAAAAAG TGGAGGAATC TAAATTAAAA 3060
CCAAATACAC GTTATATTGT TTCTGGATTC ATCGCACATG GAAAAGACCT AGAAATTGTT 3120
GTTTCACGTT ATGGGCAAGA AGTGCAAAAG GTCGTGCAAG TTCCTTATGG AGAAGCATTC 3180
CCGTTAACAT CAAATGGACC AGTTTGTTGT CCCCCACGTT CTACAAGTAA TGGAACCTTA 3240

```
GGAGATCCAC ATTTCTTTAG TTACAGTATC GATGTAGGTG CACTAGATTT ACAAGCAAAC   3300
CCTGGTATTG AATTTGGTCT TCGTATTGTA AATCCAACTG GAATGGCACG CGTAAGCAAT   3360
TTGGAAATTC GTGAAGATCG TCCATTAGCA GCAAATGAAA TACGACAAGT ACAACGTGTC   3420
GCAAGAAATT GGAGAACCGA GTATGAGAAA GAACGTGCGG AAGTAACAAG TTTAATTCAA   3480
CCTGTTATCA ATCGAATCAA CGGATTGTAT GAAAATGGAA ATTGGAACGG TTCTATTCGT   3540
TCAGATATTT CGTATCAGAA TATAGACGCG ATTGTATTAC CAACGTTACC AAAGTTACGC   3600
CATTGGTTTA TGTCAGATAG ATTCAGTGAA CAAGGAGATA TAATGGCTAA ATTCCAAGGT   3660
GCATTAAATC GTGCGTATGC ACAACTGGAA CAAAGTACGC TTCTGCATAA TGGTCATTTT   3720
ACAAAAGATG CAGCTAATTG GACAATAGAA GGCGATGCAC ATCAGATAAC ACTAGAAGAT   3780
GGTAGACGTG TATTGCGACT TCCAGATTGG TCTTCGAGTG TATCTCAAAT GATTGAAATC   3840
GAGAATTTTA ATCCAGATAA AGAATACAAC TTAGTATTCC ATGGGCAAGG AGAAGGAACG   3900
GTTACGTTGG AGCATGGAGA AGAAACAAAA TATATAGAAA CGCATACACA TCATTTTGCG   3960
AATTTTACAA CTTCTCAACG TCAAGGACTC ACGTTTGAAT CAAATAAAGT GACAGTGACC   4020
ATTTCTTCAG AAGATGGAGA ATTCTTAGTG GATAATATTG CGCTTGTGGA AGCTCCTCTT   4080
CCTACAGATG ACCAAAATTC TGAGGGAAAT ACGGCTTCCA GTACGAATAG CGATACAAGT   4140
ATGAACAACA ATCAA                                                    4155
```

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1385 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: YES

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: BACILLUS THURINGIENSIS
(C) INDIVIDUAL ISOLATE: PS17

(vii) IMMEDIATE SOURCE:
(B) CLONE: E. coli NM522(pMYC 1627) NRRL B-18651

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

```
Met Ala Ile Leu Asn Glu Leu Tyr Pro Ser Val Pro Tyr Asn Val Leu
1               5                  10                  15

Ala Tyr Thr Pro Pro Ser Phe Leu Pro Asp Ala Gly Thr Gln Ala Thr
            20                  25                  30

Pro Ala Asp Leu Thr Ala Tyr Glu Gln Leu Leu Lys Asn Leu Glu Lys
        35                  40                  45

Gly Ile Asn Ala Gly Thr Tyr Ser Lys Ala Ile Ala Asp Val Leu Lys
    50                  55                  60

Gly Ile Phe Ile Asp Asp Thr Ile Asn Tyr Gln Thr Tyr Val Asn Ile
65                  70                  75                  80

Gly Leu Ser Leu Ile Thr Leu Ala Val Pro Glu Ile Gly Ile Phe Thr
                85                  90                  95

Pro Phe Ile Gly Leu Phe Phe Ala Ala Leu Asn Lys His Asp Ala Pro
            100                 105                 110

Pro Pro Pro Asn Ala Lys Asp Ile Phe Glu Ala Met Lys Pro Ala Ile
        115                 120                 125

Gln Glu Met Ile Asp Arg Thr Leu Thr Ala Asp Glu Gln Thr Phe Leu
    130                 135                 140

Asn Gly Glu Ile Ser  ly Leu  ln Asn Leu Ala Ala Arg Tyr Gln Ser
145                 150                 155                 160
```

Thr Met Asp Asp Ile 1n Ser His Gly Gly Phe Asn Lys Val Asp Ser
165 170 175

Gly Leu Ile Lys Lys Phe Thr Asp Glu Val Leu Ser Leu Asn Ser Phe
180 185 190

Tyr Thr Asp Arg Leu Pro Val Phe Ile Thr Asp Asn Thr Ala Asp Arg
195 200 205

Thr Leu Leu Gly Leu Pro Tyr Tyr Ala Ile Leu Ala Ser Met His Leu
210 215 220

Met Leu Leu Arg Asp Ile Ile Thr Lys Gly Pro Thr Trp Asp Ser Lys
225 230 235 240

Ile Asn Phe Thr Pro Asp Ala Ile Asp Ser Phe Lys Thr Asp Ile Lys
245 250 255

Asn Asn Ile Lys Leu Tyr Ser Lys Thr Ile Tyr Asp Val Phe Gln Lys
260 265 270

Gly Leu Ala Ser Tyr Gly Thr Pro Ser Asp Leu Glu Ser Phe Ala Lys
275 280 285

Lys Gln Lys Tyr Ile Glu Ile Met Thr Thr His Cys Leu Asp Phe Ala
290 295 300

Arg Leu Phe Pro Thr Phe Asp Pro Asp Leu Tyr Pro Thr Gly Ser Gly
305 310 315 320

Asp Ile Ser Leu Gln Lys Thr Arg Arg Ile Leu Ser Pro Phe Ile Pro
325 330 335

Ile Arg Thr Ala Asp Gly Leu Thr Leu Asn Asn Thr Ser Ile Asp Thr
340 345 350

Ser Asn Trp Pro Asn Tyr Glu Asn Gly Asn Gly Ala Phe Pro Asn Pro
355 360 365

Lys Glu Arg Ile Leu Lys Gln Phe Lys Leu Tyr Pro Ser Trp Arg Ala
370 375 380

Gly Gln Tyr Gly Gly Leu Leu Gln Pro Tyr Leu Trp Ala Ile Glu Val
385 390 395 400

Gln Asp Ser Val Glu Thr Arg Leu Tyr Gly Gln Leu Pro Ala Val Asp
405 410 415

Pro Gln Ala Gly Pro Asn Tyr Val Ser Ile Asp Ser Ser Asn Pro Ile
420 425 430

Ile Gln Ile Asn Met Asp Thr Trp Lys Thr Pro Pro Gln Gly Ala Ser
435 440 445

Gly Trp Asn Thr Asn Leu Met Arg Gly Ser Val Ser Gly Leu Ser Phe
450 455 460

Leu Gln Arg Asp Gly Thr Arg Leu Ser Ala Gly Met Gly Gly Gly Phe
465 470 475 480

Ala Asp Thr Ile Tyr Ser Leu Pro Ala Thr His Tyr Leu Ser Tyr Leu
485 490 495

Tyr Gly Thr Pro Tyr Gln Thr Ser Asp Asn Tyr Ser Gly His Val Gly
500 505 510

Ala Leu Val Gly Val Ser Thr Pro Gln Glu Ala Thr Leu Pro Asn Ile
515 520 525

Ile Gly Gln Pro Asp Glu Gln Gly Asn Val Ser Thr Met Gly Phe Pro
530 535 540

Phe Glu Lys Ala Ser Tyr Gly Gly Thr Val Val Lys Glu Trp Leu Asn
545 550 555 560

Gly Ala Asn Ala Met Lys Leu Ser Pro Gly Gln Ser Ile Gly Ile Pro
565 570 575

Ile Thr Asn Val Thr Ser Gly Glu Tyr Gln Ile Arg Cys Arg Tyr Ala
580 585 590

Ser Asn Asp Asn Thr Asn Val Phe Phe Asn Val Asp Thr Gly Gly Ala
595 600 605

Asn Pro Ile Phe 1n 1n Ile Asn Phe Ala Ser Thr Val Asp Asn Asn
610 615 620

Thr Gly Val Gln Gly Ala Asn Gly Val Tyr Val Val Lys Ser Ile Ala
625 630 635 640

Thr Thr Asp Asn Ser Phe Thr Glu Ile Pro Ala Lys Thr Ile Asn Val
645 650 655

His Leu Thr Asn Gln Gly Ser Ser Asp Val Phe Leu Asp Arg Ile Glu
660 665 670

Phe Ile Pro Phe Ser Leu Pro Leu Ile Tyr His Gly Ser Tyr Asn Thr
675 680 685

Ser Ser Gly Ala Asp Asp Val Leu Trp Ser Ser Ser Asn Met Asn Tyr
690 695 700

Tyr Asp Ile Ile Val Asn Gly Gln Ala Asn Ser Ser Ser Ile Ala Ser
705 710 715 720

Ser Met His Leu Leu Asn Lys Gly Lys Val Ile Lys Thr Ile Asp Ile
725 730 735

Pro Gly His Ser Glu Thr Phe Phe Ala Thr Phe Pro Val Pro Glu Gly
740 745 750

Phe Asn Glu Val Arg Ile Leu Ala Gly Leu Pro Glu Val Ser Gly Asn
755 760 765

Ile Thr Val Gln Ser Asn Asn Pro Pro Gln Pro Ser Asn Asn Gly Gly
770 775 780

Gly Asp Gly Gly Gly Asn Gly Gly Gly Asp Gly Gly Gln Tyr Asn Phe
785 790 795 800

Ser Leu Ser Gly Ser Asp His Thr Thr Ile Tyr His Gly Lys Leu Glu
805 810 815

Thr Gly Ile His Val Gln Gly Asn Tyr Thr Tyr Thr Gly Thr Pro Val
820 825 830

Leu Ile Leu Asn Ala Tyr Arg Asn Asn Thr Val Val Ser Ser Ile Pro
835 840 845

Val Tyr Ser Pro Phe Asp Ile Thr Ile Gln Thr Glu Ala Asp Ser Leu
850 855 860

Glu Leu Glu Leu Gln Pro Arg Tyr Gly Phe Ala Thr Val Asn Gly Thr
865 870 875 880

Ala Thr Val Lys Ser Pro Asn Val Asn Tyr Asp Arg Ser Phe Lys Leu
885 890 895

Pro Ile Asp Leu Gln Asn Ile Thr Thr Gln Val Asn Ala Leu Phe Ala
900 905 910

Ser Gly Thr Gln Asn Met Leu Ala His Asn Val Ser Asp His Asp Ile
915 920 925

Glu Glu Val Val Leu Lys Val Asp Ala Leu Ser Asp Glu Val Phe Gly
930 935 940

Asp Glu Lys Lys Ala Leu Arg Lys Leu Val Asn Gln Ala Lys Arg Leu
945 950 955 960

Ser Arg Ala Arg Asn Leu Leu Ile Gly Gly Ser Phe Glu Asn Trp Asp
965 970 975

Ala Trp Tyr Lys Gly Arg Asn Val Val Thr Val Ser Asp His Glu Leu
980 985 990

Phe Lys Ser Asp His Val Leu Leu Pro Pro Pro Gly Leu Ser Pro Ser
995 1000 1005

Tyr Ile Phe Gln Lys Val Glu Glu Ser Lys Leu Lys Pro Asn Thr Arg
1010 1015 1020

Tyr Ile Val Ser Gly Phe Ile Ala His Gly Lys Asp Leu Glu Ile Val
1025 1030 1035 1040

Val Ser Arg Tyr Gly Gln Glu Val Gln Lys Val Val Gln Val Pro Tyr
1045 1050 1055

Gly lu Ala Phe Pro Leu Thr Ser Asn ly Pro Val Cys Cys Pro Pro
1060 1065 1070

Arg Ser Thr Ser Asn Gly Thr Leu Gly Asp Pro His Phe Ph s r Tyr
1075 1080 1085

Ser Ile Asp Val Gly Ala Leu Asp Leu Gln Ala Asn Pro Gly Ile Glu
1090 1095 1100

Phe Gly Leu Arg Ile Val Asn Pro Thr Gly Met Ala Arg Val Ser Asn
1105 1110 1115 1120

Leu Glu Ile Arg Glu Asp Arg Pro Leu Ala Ala Asn Glu Ile Arg Gln
1125 1130 1135

Val Gln Arg Val Ala Arg Asn Trp Arg Thr Glu Tyr Glu Lys Glu Arg
1140 1145 1150

Ala Glu Val Thr Ser Leu Ile Gln Pro Val Ile Asn Arg Ile Asn Gly
1155 1160 1165

Leu Tyr Glu Asn Gly Asn Trp Asn Gly Ser Ile Arg Ser Asp Ile Ser
1170 1175 1180

Tyr Gln Asn Ile Asp Ala Ile Val Leu Pro Thr Leu Pro Lys Leu Arg
1185 1190 1195 1200

His Trp Phe Met Ser Asp Arg Phe Ser Glu Gln Gly Asp Ile Met Ala
1205 1210 1215

Lys Phe Gln Gly Ala Leu Asn Arg Ala Tyr Ala Gln Leu Glu Gln Ser
1220 1225 1230

Thr Leu Leu His Asn Gly His Phe Thr Lys Asp Ala Ala Asn Trp Thr
1235 1240 1245

Ile Glu Gly Asp Ala His Gln Ile Thr Leu Glu Asp Gly Arg Arg Val
1250 1255 1260

Leu Arg Leu Pro Asp Trp Ser Ser Ser Val Ser Gln Met Ile Glu Ile
1265 1270 1275 1280

Glu Asn Phe Asn Pro Asp Lys Glu Tyr Asn Leu Val Phe His Gly Gln
1285 1290 1295

Gly Glu Gly Thr Val Thr Leu Glu His Gly Glu Glu Thr Lys Tyr Ile
1300 1305 1310

Glu Thr His Thr His His Phe Ala Asn Phe Thr Thr Ser Gln Arg Gln
1315 1320 1325

Gly Leu Thr Phe Glu Ser Asn Lys Val Thr Val Thr Ile Ser Ser Glu
1330 1335 1340

Asp Gly Glu Phe Leu Val Asp Asn Ile Ala Leu Val Glu Ala Pro Leu
1345 1350 1355 1360

Pro Thr Asp Asp Gln Asn Ser Glu Gly Asn Thr Ala Ser Ser Thr Asn
1365 1370 1375

Ser Asp Thr Ser Met Asn Asn Asn Gln
1380 1385

(2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 3867 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: Bacillus thuringiensis
(B) STRAIN: PS17
(C) INDIVIDUAL ISOLATE: PS17b

(vii) IMMEDIATE SOURCE:
(B) CLONE: E. coli NM522(pMYC 1628) NRRL B-18652

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

ATGGCAATTT TAAATGAATT ATATCCATCT GTACCTTATA ATGTATTGGC GTATACGCCA 60

CCCTCTTTTT TACCTGATGC GGGTACACAA GCTACACCTG CTGACTTAAC AGCTTATGAA 120

```
CAATTGTTGA AAAATTTAGA AAAAGGGATA AATGCTGGAA CTTATTCGAA AGCAATAGCT   180
GATGTACTTA AAGGTATTTT TATAGATGAT ACAATAAATT ATCAAACATA TGTAAATATT   240
GGTTTAAGTT TAATTACATT AGCTGTACCG GAAATTGGTA TTTTTACACC TTTCATCGGT   300
TTGTTTTTTG CTGCATTGAA TAAACATGAT GCTCCACCTC CTCCTAATGC AAAAGATATA   360
TTTGAGGCTA TGAAACCAGC GATTCAAGAG ATGATTGATA GAACTTTAAC TGCGGATGAG   420
CAAACATTTT TAAATGGGGA AATAAGTGGT TTACAAAATT TAGCAGCAAG ATACCAGTCT   480
ACAATGGATG ATATTCAAAG CCATGGAGGA TTTAATAAGG TAGATTCTGG ATTAATTAAA   540
AAGTTTACAG ATGAGGTACT ATCTTTAAAT AGTTTTTATA CAGATCGTTT ACCTGTATTT   600
ATTACAGATA ATACAGCGGA TCGAACTTTG TTAGGTCTTC CTTATTATGC TATACTTGCG   660
AGCATGCATC TTATGTTATT AAGAGATATC ATTACTAAGG GTCCGACATG GGATTCTAAA   720
ATTAATTTCA CACCAGATGC AATTGATTCC TTTAAAACCG ATATTAAAAA TAATATAAAG   780
CTTTACTCTA AAACTATTTA TGACGTATTT CAGAAGGGAC TTGCTTCATA CGGAACGCCT   840
TCTGATTTAG AGTCCTTTGC AAAAAAACAA AAATATATTG AAATTATGAC AACACATTGT   900
TTAGATTTTG CAAGATTGTT TCCTACTTTT GATCCAGATC TTTATCCAAC AGGATCAGGT   960
GATATAAGTT TACAAAAAAC ACGTAGAATT CTTTCTCCTT TTATCCCTAT ACGTACTGCA  1020
GATGGGTTAA CATTAAATAA TACTTCAATT GATACTTCAA ATTGGCCTAA TTATGAAAAT  1080
GGGAATGGCG CGTTTCCAAA CCCAAAAGAA AGAATATTAA AACAATTCAA ACTGTATCCT  1140
AGTTGGAGAG CGGCACAGTA CGGTGGGCTT TTACAACCTT ATTTATGGGC AATAGAAGTC  1200
CAAGATTCTG TAGAGACTCG TTTGTATGGG CAGCTTCCAG CTGTAGATCC ACAGGCAGGG  1260
CCTAATTATG TTTCCATAGA TTCTTCTAAT CCAATCATAC AAATAAATAT GGATACTTGG  1320
AAAACACCAC CACAAGGTGC GAGTGGGTGG AATACAAATT TAATGAGAGG AAGTGTAAGC  1380
GGGTTAAGTT TTTTACAACG AGATGGTACG AGACTTAGTG CTGGTATGGG TGGTGGTTTT  1440
GCTGATACAA TATATAGTCT CCCTGCAACT CATTATCTTT CTTATCTCTA TGGAACTCCT  1500
TATCAAACTT CTGATAACTA TTCTGGTCAC GTTGGTGCAT TGGTAGGTGT GAGTACGCCT  1560
CAAGAGGCTA CTCTTCCTAA TATTATAGGT CAACCAGATG AACAGGGAAA TGTATCTACA  1620
ATGGGATTTC CGTTTGAAAA AGCTTCTTAT GGAGGTACAG TTGTTAAAGA ATGGTTAAAT  1680
GGTGCGAATG CGATGAAGCT TTCTCCTGGG CAATCTATAG GTATTCCTAT TACAAATGTA  1740
ACAAGTGGAG AATATCAAAT TCGTTGTCGT TATGCAAGTA ATGATAATAC TAACGTTTTC  1800
TTTAATGTAG ATACTGGTGG AGCAAATCCA ATTTTCCAAC AGATAAACTT TGCATCTACT  1860
GTAGATAATA ATACGGGAGT ACAAGGAGCA AATGGTGTCT ATGTAGTCAA ATCTATTGCT  1920
ACAACTGATA ATTCTTTTAC AGTAAAAATT CCTGCGAAGA CGATTAATGT TCATTTAACC  1980
AACCAAGGTT CTTCTGATGT CTTTTTAGAT CGTATTGAGT TTGTTCCAAT TCTAGAATCA  2040
AATACTGTAA CTATATTCAA CAATTCATAT ACTACAGGTT CAGCAAATCT TATACCAGCA  2100
ATAGCTCCTC TTTGGAGTAC TAGTTCAGAT AAAGCCCTTA CAGGTTCTAT GTCAATAACA  2160
GGTCGAACTA CCCCTAACAG TGATGATGCT TTGCTTCGAT TTTTTAAAAC TAATTATGAT  2220
ACACAAACCA TTCCTATTCC GGGTTCCGGA AAAGATTTTA CAAATACTCT AGAAATACAA  2280
GACATAGTTT CTATTGATAT TTTTGTCGGA TCTGGTCTAC ATGGATCCGA TGGATCTATA  2340
AAATTAGATT TTACCAATAA TAATAGTGGT AGTGGTGGCT CTCCAAAGAG TTTCACCGAG  2400
CAAATTGATT TAGAGAATAT CACAACACAA GTGAATGCTC TATTCACATC TAATACACAA  2460
GATGCACTTG CAACAGATGT GAGTGATCAT GATATTGAAG AAGTGGTTCT AAAAGTAGAT  2520
GCATTATCTG ATGAAGTGTT TGGAAAAGAG AAAAAAACAT TGCGTAAATT TGTAAATCAA  2580
GCGAAGCGCT TAAGCAAGGC GCGTAATCTC CTGGTAGGAG GCAATTTTGA TAACTTGGAT  2640
GCTTGGTATA GAGGAAGAAA TGTAGTAAAC GTATCTAATC ACGAACTGTT GAAGAGTGAT  2700
CATGTATTAT TACCACCACC AGGATTGTCT CCATCTTATA TTTTCCAAAA AGTGGAGGAA  2760
```

```
TCTAAATTAA AACGAAATAC ACGTTATACG GTTTCTGGAT TTATTGCGCA TGCAACAGAT   2820
TTAGAAATTG TGGTTTCTCG TTATGGGCAA GAAATAAAGA AAGTGGTGCA AGTTCCTTAT   2880
GGAGAAGCAT TCCCATTAAC ATCAAGTGGA CCAGTTTGTT GTATCCCACA TTCTACAAGT   2940
AATGGAACTT TAGGCAATCC ACATTTCTTT AGTTACAGTA TTGATGTAGG TGCATTAGAT   3000
GTAGACACAA ACCCTGGTAT TGAATTCGGT CTTCGTATTG TAAATCCAAC TGGAATGGCA   3060
CGCGTAAGCA ATTTGGAAAT TCGTGAAGAT CGTCCATTAG CAGCAAATGA AATACGACAA   3120
GTACAACGTG TCGCAAGAAA TTGGAGAACC GAGTATGAGA AAGAACGTGC GGAAGTAACA   3180
AGTTTAATTC AACCTGTTAT CAATCGAATC AATGGATTGT ATGACAATGG AAATTGGAAC   3240
GGTTCTATTC GTTCAGATAT TTCGTATCAG AATATAGACG CGATTGTATT ACCAACGTTA   3300
CCAAAGTTAC GCCATTGGTT TATGTCAGAT AGATTTAGTG AACAAGGAGA TATCATGGCT   3360
AAATTCCAAG GTGCATTAAA TCGTGCGTAT GCACAACTGG AACAAAATAC GCTTCTGCAT   3420
AATGGTCATT TTACAAAAGA TGCAGCCAAT TGGACGGTAG AAGGCGATGC ACATCAGGTA   3480
GTATTAGAAG ATGGTAAACG TGTATTACGA TTGCCAGATT GGTCTTCGAG TGTGTCTCAA   3540
ACGATTGAAA TCGAGAATTT TGATCCAGAT AAAGAATATC AATTAGTATT TCATGGGCAA   3600
GGAGAAGGAA CGGTTACGTT GGAGCATGGA GAAGAAACAA AATATATAGA AACGCATACA   3660
CATCATTTTG CGAATTTTAC AACTTCTCAA CGTCAAGGAC TCACGTTTGA ATCAAATAAA   3720
GTGACAGTGA CCATTTCTTC AGAAGATGGA GAATTCTTAG TGGATAATAT TGCGCTTGTG   3780
GAAGCTCCTC TTCCTACAGA TGACCAAAAT TCTGAGGGAA ATACGGCTTC CAGTACGAAT   3840
AGCGATACAA GTATGAACAA CAATCAA                                       3867
```

(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1289 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: YES

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: BACILLUS THURINGIENSIS
(C) INDIVIDUAL ISOLATE: PS17

(vii) IMMEDIATE SOURCE:
(B) CLONE: E. coli NM522(pMYC 1628) NRRL B-18652

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

```
Met Ala Ile Leu Asn Glu Leu Tyr Pro Ser Val Pro Tyr Asn Val Leu
1               5                  10                  15

Ala Tyr Thr Pro Pro Ser Phe Leu Pro Asp Ala Gly Thr Gln Ala Thr
            20                  25                  30

Pro Ala Asp Leu Thr Ala Tyr Glu Gln Leu Leu Lys Asn Leu Glu Lys
        35                  40                  45

Gly Ile Asn Ala Gly Thr Tyr Ser Lys Ala Ile Ala Asp Val Leu Lys
    50                  55                  60

Gly Ile Phe Ile Asp Asp Thr Ile Asn Tyr Gln Thr Tyr Val Asn Ile
65                  70                  75                  80

Gly Leu Ser Leu Ile Thr Leu Ala Val Pro Glu Ile Gly Ile Phe Thr
                85                  90                  95

Pro Phe Il  Gly Leu Phe Phe Ala Ala Leu Asn Lys His Asp Ala Pro
            100                 105                 110

Pro Pro Pro Asn Ala Lys Asp Ile Phe Glu Ala Met Lys Pro Ala Ile
        115                 120                 125
```

Gln Glu Met Ile Asp Arg Thr Leu Thr Ala Asp Glu Gln Thr Phe Leu
130 135 140

Asn Gly Glu Ile Ser Gly Leu Gln Asn Leu Ala Ala Arg Tyr Gln Ser
145 150 155 160

Thr Met Asp Asp Ile Gln Ser His Gly Gly Phe Asn Lys Val Asp Ser
165 170 175

Gly Leu Ile Lys Lys Phe Thr Asp Glu Val Leu Ser Leu Asn Ser Phe
180 185 190

Tyr Thr Asp Arg Leu Pro Val Phe Ile Thr Asp Asn Thr Ala Asp Arg
195 200 205

Thr Leu Leu Gly Leu Pro Tyr Tyr Ala Ile Leu Ala Ser Met His Leu
210 215 220

Met Leu Leu Arg Asp Ile Ile Thr Lys Gly Pro Thr Trp Asp Ser Lys
225 230 235 240

Ile Asn Phe Thr Pro Asp Ala Ile Asp Ser Phe Lys Thr Asp Ile Lys
245 250 255

Asn Asn Ile Lys Leu Tyr Ser Lys Thr Ile Tyr Asp Val Phe Gln Lys
260 265 270

Gly Leu Ala Ser Tyr Gly Thr Pro Ser Asp Leu Glu Ser Phe Ala Lys
275 280 285

Lys Gln Lys Tyr Ile Glu Ile Met Thr Thr His Cys Leu Asp Phe Ala
290 295 300

Arg Leu Phe Pro Thr Phe Asp Pro Asp Leu Tyr Pro Thr Gly Ser Gly
305 310 315 320

Asp Ile Ser Leu Gln Lys Thr Arg Arg Ile Leu Ser Pro Phe Ile Pro
325 330 335

Ile Arg Thr Ala Asp Gly Leu Thr Leu Asn Asn Thr Ser Ile Asp Thr
340 345 350

Ser Asn Trp Pro Asn Tyr Glu Asn Gly Asn Gly Ala Phe Pro Asn Pro
355 360 365

Lys Glu Arg Ile Leu Lys Gln Phe Lys Leu Tyr Pro Ser Trp Arg Ala
370 375 380

Ala Gln Tyr Gly Gly Leu Gln Pro Tyr Leu Trp Ala Ile Glu Val
385 390 395 400

Gln Asp Ser Val Glu Thr Arg Leu Tyr Gly Gln Leu Pro Ala Val Asp
405 410 415

Pro Gln Ala Gly Pro Asn Tyr Val Ser Ile Asp Ser Ser Asn Pro Ile
420 425 430

Ile Gln Ile Asn Met Asp Thr Trp Lys Thr Pro Pro Gln Gly Ala Ser
435 440 445

Gly Trp Asn Thr Asn Leu Met Arg Gly Ser Val Ser Gly Leu Ser Phe
450 455 460

Leu Gln Arg Asp Gly Thr Arg Leu Ser Ala Gly Met Gly Gly Gly Phe
465 470 475 480

Ala Asp Thr Ile Tyr Ser Leu Pro Ala Thr His Tyr Leu Ser Tyr Leu
485 490 495

Tyr Gly Thr Pro Tyr Gln Thr Ser Asp Asn Tyr Ser Gly His Val Gly
500 505 510

Ala Leu Val Gly Val Ser Thr Pro Gln Glu Ala Thr Leu Pro Asn Ile
515 520 525

Ile Gly Gln Pro Asp Glu Gln Gly Asn Val Ser Thr Met Gly Phe Pro
530 535 540

Phe Glu Lys Ala Ser Tyr Gly Gly Thr Val Val Lys Glu Trp Leu Asn
545 550 555 560

Gly Ala Asn Ala Met Lys Leu Ser Pro Gly Gln Ser Ile Gly Ile Pro
565 570 575

Ile Thr Asn Val Thr Ser Gly Glu Tyr Gln Ile Arg Cys Arg Tyr Ala
580 585 590

Ser Asn Asp Asn Thr Asn Val Phe Phe Asn Val Asp Thr Gly ly Ala
595 600 605

Asn Pro Ile Phe Gln Gln Ile Asn Phe Ala Ser Thr Val Asp Asn Asn
610 615 620

Thr Gly Val Gln Gly Ala Asn Gly Val Tyr Val Val Lys Ser Ile Ala
625 630 635 640

Thr Thr Asp Asn Ser Phe Thr Val Lys Ile Pro Ala Lys Thr Ile Asn
645 650 655

Val His Leu Thr Asn Gln Gly Ser Ser Asp Val Phe Leu Asp Arg Ile
660 665 670

Glu Phe Val Pro Ile Leu Glu Ser Asn Thr Val Thr Ile Phe Asn Asn
675 680 685

Ser Tyr Thr Thr Gly Ser Ala Asn Leu Ile Pro Ala Ile Ala Pro Leu
690 695 700

Trp Ser Thr Ser Ser Asp Lys Ala Leu Thr Gly Ser Met Ser Ile Thr
705 710 715 720

Gly Arg Thr Thr Pro Asn Ser Asp Asp Ala Leu Leu Arg Phe Phe Lys
725 730 735

Thr Asn Tyr Asp Thr Gln Thr Ile Pro Ile Pro Gly Ser Gly Lys Asp
740 745 750

Phe Thr Asn Thr Leu Glu Ile Gln Asp Ile Val Ser Ile Asp Ile Phe
755 760 765

Val Gly Ser Gly Leu His Gly Ser Asp Gly Ser Ile Lys Leu Asp Phe
770 775 780

Thr Asn Asn Asn Ser Gly Ser Gly Gly Ser Pro Lys Ser Phe Thr Glu
785 790 795 800

Gln Asn Asp Leu Glu Asn Ile Thr Thr Gln Val Asn Ala Leu Phe Thr
805 810 815

Ser Asn Thr Gln Asp Ala Leu Ala Thr Asp Val Ser Asp His Asp Ile
820 825 830

Glu Glu Val Val Leu Lys Val Asp Ala Leu Ser Asp Glu Val Phe Gly
835 840 845

Lys Glu Lys Lys Thr Leu Arg Lys Phe Val Asn Gln Ala Lys Arg Leu
850 855 860

Ser Lys Ala Arg Asn Leu Leu Val Gly Gly Asn Phe Asp Asn Leu Asp
865 870 875 880

Ala Trp Tyr Arg Gly Arg Asn Val Val Asn Val Ser Asn His Glu Leu
885 890 895

Leu Lys Ser Asp His Val Leu Leu Pro Pro Pro Gly Leu Ser Pro Ser
900 905 910

Tyr Ile Phe Gln Lys Val Glu Glu Ser Lys Leu Lys Arg Asn Thr Arg
915 920 925

Tyr Thr Val Ser Gly Phe Ile Ala His Ala Thr Asp Leu Glu Ile Val
930 935 940

Val Ser Arg Tyr Gly Gln Glu Ile Lys Lys Val Val Gln Val Pro Tyr
945 950 955 960

Gly Glu Ala Phe Pro Leu Thr Ser Ser Gly Pro Val Cys Cys Ile Pro
965 970 975

His Ser Thr Ser Asn Gly Thr Leu Gly Asn Pro His Phe Phe Ser Tyr
980 985 990

Ser Ile Asp Val Gly Ala Leu Asp Val Asp Thr Asn Pro Gly Ile Glu
995 1000 1005

Phe Gly Leu Arg Ile Val Asn Pro Thr Gly Met Ala Arg Val Ser Asn
1010 1015 1020

Leu Glu Ile Arg lu Asp Arg Pro Leu Ala Ala Asn Glu Ile Arg Gln
1025 1030 1035 1040

Val Gln Arg Val Ala Arg Asn Trp Arg Thr Glu Tyr Glu Lys Glu Arg
1045 1050 1055

Ala Glu Val Thr Ser Leu Ile Gln Pro Val Ile Asn Arg Ile Asn Gly
1060 1065 1070

Leu Tyr Asp Asn Gly Asn Trp Asn Gly Ser Ile Arg Ser Asp Ile Ser
1075 1080 1085

Tyr Gln Asn Ile Asp Ala Ile Val Leu Pro Thr Leu Pro Lys Leu Arg
1090 1095 1100

His Trp Phe Met Ser Asp Arg Phe Ser Glu Gln Gly Asp Ile Met Ala
1105 1110 1115 1120

Lys Phe Gln Gly Ala Leu Asn Arg Ala Tyr Ala Gln Leu Glu Gln Asn
1125 1130 1135

Thr Leu Leu His Asn Gly His Phe Thr Lys Asp Ala Ala Asn Trp Thr
1140 1145 1150

Val Glu Gly Asp Ala His Gln Val Val Leu Glu Asp Gly Lys Arg Val
1155 1160 1165

Leu Arg Leu Pro Asp Trp Ser Ser Ser Val Ser Gln Thr Ile Glu Ile
1170 1175 1180

Glu Asn Phe Asp Pro Asp Lys Glu Tyr Gln Leu Val Phe His Gly Gln
1185 1190 1195 1200

Gly Glu Gly Thr Val Thr Leu Glu His Gly Glu Glu Thr Lys Tyr Ile
1205 1210 1215

Glu Thr His Thr His His Phe Ala Asn Phe Thr Thr Ser Gln Arg Gln
1220 1225 1230

Gly Leu Thr Phe Glu Ser Asn Lys Val Thr Val Thr Ile Ser Ser Glu
1235 1240 1245

Asp Gly Glu Phe Leu Val Asp Asn Ile Ala Leu Val Glu Ala Pro Leu
1250 1255 1260

Pro Thr Asp Asp Gln Asn Ser Glu Gly Asn Thr Ala Ser Ser Thr Asn
1265 1270 1275 1280

Ser Asp Thr Ser Met Asn Asn Asn Gln
1285

(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 3771 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: Bacillus thuringiensis
(C) INDIVIDUAL ISOLATE: 33f2

(vii) IMMEDIATE SOURCE:
(B) CLONE: E. coli NM522(pMYC 2316) B-18785

(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 4..24
(D) OTHER INFORMATION: /function= "oligonucleotide hybridization probe"
/product= "GCA/T ACA/T TTA AAT GAA GTA/T TAT"
/standard_name= "probe a"
/note= "Probe A"

(ix) FEATURE:
(A) NAME/KEY: misc_feature
(B) LOCATION: 13..33
(D) OTHER INFORMATION: /function= "oligonucleotide hybridization probe"
/product= "AAT GAA GTA/T TAT CCA/T GTA/T AAT"
/standard_name= "Probe B"
/lab_l= probe-b
/note= "probe b"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

ATGGCTACAC TTAATGAAGT ATATCCTGTG AATTATAATG TATTATCTTC TGATGCTTTT 60
CAACAATTAG ATACAACAGG TTTTAAAAGT AAATATGATG AAATGATAAA AGCATTCGAA 120
AAAAAATGGA AAAAAGGGGC AAAAGGAAAA GACCTTTTAG ATGTTGCATG GACTTATATA 180
ACTACAGGAG AAATTGACCC TTTAAATGTA ATTAAAGGTG TTTTATCTGTATTAACTTTA 240
ATTCCTGAAG TTGGTACTGT GGCCTCTGCA GCAAGTACTA TTGTAAGTTT TATTTGGCCT 300
AAAATATTTG GAGATAAACC AAATGCAAAA AATATATTTG AAGAGCTCAA GCCTCAAATT 360
GAAGCATTAA TTCAACAAGA TATAACAAAC TATCAAGATG CAATTAATCA AAAAAAATTT 420
GACAGTCTTC AGAAAACAAT TAATCTATAT ACAGTAGCTA TAGATAACAA TGATTACGTA 480
ACAGCAAAAA CGCAACTCGA AAATCTAAAT TCTATACTTA CCTCAGATAT CTCCATATTT 540
ATTCCAGAAG GATATGAAAC TGGAGGTTTA CCTTATTATG CTATGGTTGC TAATGCTCAT 600
ATATTATTGT TAAGAGACGC TATAGTTAAT GCAGAGAAAT TAGGCTTTAG TGATAAAGAA 660
GTAGACACAC ATAAAAAATA TATCAAAATG ACAATACACA ATCATACTGA AGCAGTAATA 720
AAAGCATTCT TAAATGGACT TGACAAATTT AAGAGTTTAG ATGTAAATAG CTATAATAAA 780
AAAGCAAATT ATATTAAAGG TATGACAGAA ATGGTTCTTG ATCTAGTTGC TCTATGGCCA 840
ACTTTCGATC CAGATCATTA TCAAAAAGAA GTAGAAATTG AATTTACAAG AACTATTTCT 900
TCTCCAATTT ACCAACCTGT ACCTAAAAAC ATGCAAAATA CCTCTAGCTC TATTGTACCT 960
AGCGATCTAT TTCACTATCA AGGAGATCTT GTAAAATTAG AATTTTCTAC AAGAACGGAC 1020
AACGATGGTC TTGCAAAAAT TTTTACTGGT ATTCGAAACA CATTCTACAA ATCGCCTAAT 1080
ACTCATGAAA CATACCATGT AGATTTTAGT TATAATACCC AATCTAGTGG TAATATTTCA 1140
AGAGGCTCTT CAAATCCGAT TCCAATTGAT CTTAATAATC CCATTATTTC AACTTGTATT 1200
AGAAATTCAT TTTATAAGGC AATAGCGGGA TCTTCTGTTT TAGTTAATTT TAAAGATGGC 1260
ACTCAAGGGT ATGCATTTGC CCAAGCACCA ACAGGAGGTG CCTGGGACCA TTCTTTTATT 1320
GAATCTGATG GTGCCCCAGA AGGGCATAAA TTAAACTATA TTTATACTTC TCCAGGTGAT 1380
ACATTAAGAG ATTTCATCAA TGTATATACT CTTATAAGTA CTCCAACTAT AAATGAACTA 1440
TCAACAGAAA AAATCAAAGG CTTTCCTGCG GAAAAAGGAT ATATCAAAAA TCAAGGGATC 1500
ATGAAATATT ACGGTAAACC AGAATATATT AATGGAGCTC AACCAGTTAA TCTGGAAAAC 1560
CAGCAAACAT TAATATTCGA ATTTCATGCT TCAAAAACAG CTCAATATAC CATTCGTATA 1620
CGTTATGCCA GTACCCAAGG AACAAAAGGT TATTTTCGTT TAGATAATCA GGAACTGCAA 1680
ACGCTTAATA TACCTACTTC ACACAACGGT TATGTAACCG GTAATATTGG TGAAAATTAT 1740
GATTTATATA CAATAGGTTC ATATACAATT ACAGAAGGTA ACCATACTCT TCAAATCCAA 1800
CATAATGATA AAAATGGAAT GGTTTTAGAT CGTATTGAAT TTGTTCCTAA AGATTCACTT 1860
CAAGATTCAC CTCAAGATTC ACCTCCAGAA GTTCACGAAT CAACAATTAT TTTTGATAAA 1920
TCATCTCCAA CTATATGGTC TTCTAACAAA CACTCATATA GCCATATACA TTTAGAAGGA 1980
TCATATACAA GTCAGGGAAG TTATCCACAC AATTTATTAA TTAATTTATT TCATCCTACA 2040
GACCCTAACA GAAATCATAC TATTCATGTT AACAATGGTG ATATGAATGT TGATTATGGA 2100
AAAGATTCTG TAGCCGATGG GTTAAATTTT AATAAAATAA CTGCTACGAT ACCAAGTGAT 2160
GCTTGGTATA GCGGTACTAT TACTTCTATG CACTTATTTA ATGATAATAA TTTTAAAACA 2220
ATAACTCCTA AATTTGAACT TTCTAATGAA TTAGAAAACA TCACAACTCA AGTAAATGCT 2280
TTATTCGCAT CTAGTGCACA AGATACTCTC GCAAGTAATG TAAGTGATTA CTGGATTGAA 2340
CAGGTCGTTA TGAAAGTCGA TGCCTTATCA GATGAAGTAT TTGGAAAAGA GAAAAAAGCA 2400
TTACGTAAAT TGGTAAATCA AGCAAAACGT CTCAGTAAAA TACGAAATCT TCTCATAGGT 2460
GGTAATTTTG ACAATTTAGT CGCTTGGTAT ATGGGAAAAG ATGTAGTAAA AGAATCGGAT 2520
CATGAATTAT TTAAAAGTGA TCATGTCTTA CTACCTCCCC CAACATTCCA TCCTTCTTAT 2580

ATTTTCCAAA AGGTGGAAGA ATCAAAACTA AAACCAAATA CACGTTATAC TATTTCTGGT 2640
TTTATCGCAC ATGGAGAAGA TGTAGAGCTT TTGTCTCTC GTTATGGGCA AGAAATACAA 2700
AAAGTGATGC AAGTGCCATA TGAAGAAGCA CTTCCTCTTA CATCTGAATC TAATTCTAGT 2760
TGTTGTGTTC CAAATTTAAA TATAAATGAA ACACTAGCTG ATCCACATTT CTTTAGTTAT 2820
AGCATCGATG TTGGTTCTCT GGAAATGGAA GCGAATCCTG GTATTGAATT TGGTCTCCGT 2880
ATTGTCAAAC CAACAGGTAT GGCACGTGTA AGTAATTTAG AAATTCGAGA AGACCGTCCA 2940
TTAACAGCAA AAGAAATTCG TCAAGTACAA CGTGCAGCAA GAGATTGGAA ACAAAACTAT 3000
GAACAAGAAC GAACAGAGAT CACAGCTATA ATTCAACCTG TTCTTAATCA AATTAATGCG 3060
TTATACGAAA ATGAAGATTG GAATGGTTCT ATTCGTTCAA ATGTTTCCTA TCATGATCTA 3120
GAGCAAATTA TGCTTCCTAC TTTATTAAAA ACTGAGGAAA TAAATTGTAA TTATGATCAT 3180
CCAGCTTTTT TATTAAAAGT ATATCATTGG TTTATGACAG ATCGTATAGG AGAACATGGT 3240
ACTATTTTAG CACGTTTCCA AGAAGCATTA GATCGTGCAT ATACACAATT AGAAAGTCGT 3300
AATCTCCTGC ATAACGGTCA TTTTACAACT GATACAGCGA ATTGGACAAT AGAAGGAGAT 3360
GCCCATCATA CAATCTTAGA AGATGGTAGA CGTGTGTTAC GTTTACCAGA TTGGTCTTCT 3420
AATGCAACTC AAACAATTGA AATTGAAGAT TTTGACTTAG ATCAAGAATA CCAATTGCTC 3480
ATTCATGCAA AAGGAAAAGG TTCCATTACT TTACAACATG GAGAAGAAAA CGAATATGTG 3540
GAAACACATA CTCATCATAC AAATGATTTT ATAACATCCC AAAATATTCC TTTCACTTTT 3600
AAAGGAAATC AAATTGAAGT CCATATTACT TCAGAAGATG GAGAGTTTTT AATCGATCAC 3660
ATTACAGTAA TAGAAGTTTC TAAAACAGAC ACAAATACAA ATATTATTGA AAATTCACCA 3720
ATCAATACAA GTATGAATAG TAATGTAAGA GTAGATATAC CAAGAAGTCT C 3771

(2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1425 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: BACILLUS THURINGIENSIS
(C) INDIVIDUAL ISOLATE: PS52A1

(vii) IMMEDIATE SOURCE:
(B) CLONE: E. coli NM522(pMYC 2321) B-18770

(ix) FEATURE:
(A) NAME/KEY: mat_peptide
(B) LOCATION: 1..1425
(D) OTHER INFORMATION: /product= "OPEN READING FRAME OF MATURE PROTEIN"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

ATGATTATTG ATAGTAAAAC GACTTTACCT AGACATTCAC TTATTCATAC AATTAAATTA 60
AATTCTAATA AGAAATATGG TCCTGGTGAT ATGACTAATG GAAATCAATT TATTATTTCA 120
AAACAAGAAT GGGCTACGAT TGGAGCATAT ATTCAGACTG GATTAGGTTT ACCAGTAAAT 180
GAACAACAAT TAAGAACACA TGTTAATTTA AGTCAGGATA TATCAATACC TAGTGATTTT 240
TCTCAATTAT ATGATGTTTA TTGTTCTGAT AAAACTTCAG CAGAATGGTG GAATAAAAAT 300
TTATATCCTT TAATTATTAA ATCTGCTAAT GATATTGCTT CATATGGTTT TAAAGTTGCT 360
GGTGATCCTT CTATTAAGAA AGATGGATAT TTTAAAAAAT TGCAAGATGA ATTAGATAAT 420
ATTGTTGATA ATAATTCCGA TGATGATGCA ATAGCTAAAG CTATTAAAGA TTTTAAAGCG 480
CGATGTGGTA TTTTAATTAA AGAAGCTAAA CAATATGAAG AAGCTGCAAA AAATATTGTA 540

```
ACATCTTTAG ATCAATTTTT ACATGGTGAT CAGAAAAAAT TAGAAGGTGT TATCAATATT    600
CAAAAACGTT TAAAAGAAGT TCAAACAGCT CTTAATCAAG CCCATGGGGA AAGTAGTCCA    660
GCTCATAAAG AGTTATTAGA AAAAGTAAAA AATTTAAAAA CAACATTAGA AAGGACTATT    720
AAAGCTGAAC AAGATTTAGA GAAAAAAGTA GAATATAGTT TTCTATTAGG ACCATTGTTA    780
GGATTTGTTG TTTATGAAAT TCTTGAAAAT ACTGCTGTTC AGCATATAAA AAATCAAATT    840
GATGAGATAA AGAAACAATT AGATTCTGCT CAGCATGATT TGGATAGAGA TGTTAAAATT    900
ATAGGAATGT TAAATAGTAT TAATACAGAT ATTGATAATT TATATAGTCA AGGACAAGAA    960
GCAATTAAAG TTTTCCAAAA GTTACAAGGT ATTTGGGCTA CTATTGGAGC TCAAATAGAA   1020
AATCTTAGAA CAACGTCGTT ACAAGAAGTT CAAGATTCTG ATGATGCTGA TGAGATACAA   1080
ATTGAACTTG AGGACGCTTC TGATGCTTGG TTAGTTGTGG CTCAAGAAGC TCGTGATTTT   1140
ACACTAAATG CTTATTCAAC TAATAGTAGA CAAAATTTAC CGATTAATGT TATATCAGAT   1200
TCATGTAATT GTTCAACAAC AAATATGACA TCAAATCAAT ACAGTAATCC AACAACAAAT   1260
ATGACATCAA ATCAATATAT GATTTCACAT GAATATACAA GTTTACCAAA TAATTTTATG   1320
TTATCAAGAA ATAGTAATTT AGAATATAAA TGTCCTGAAA ATAATTTTAT GATATATTGG   1380
TATAATAATT CGGATTGGTA TAATAATTCG GATTGGTATA ATAAT                   1425
```

(2) INFORMATION FOR SEQ ID NO:7 (PS52A1):

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 475 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: YES

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: BACILLUS THURINGIENSIS
(C) INDIVIDUAL ISOLATE: PS52A1

(vii) IMMEDIATE SOURCE:
(B) CLONE: E. coli NM522(pMYC 2321) B-18770

(ix) FEATURE:
(A) NAME/KEY: Protein
(B) LOCATION: 1..475

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

```
Met Ile Ile Asp Ser Lys Thr Thr Leu Pro Arg His Ser Leu Ile His
 1               5                  10                  15

Thr Ile Lys Leu Asn Ser Asn Lys Lys Tyr Gly Pro Gly Asp Met Thr
            20                  25                  30

Asn Gly Asn Gln Phe Ile Ile Ser Lys Gln Glu Trp Ala Thr Ile Gly
        35                  40                  45

Ala Tyr Ile Gln Thr Gly Leu Gly Leu Pro Val Asn Glu Gln Gln Leu
    50                  55                  60

Arg Thr His Val Asn Leu Ser Gln Asp Ile Ser Ile Pro Ser Asp Phe
65                  70                  75                  80

Ser Gln Leu Tyr Asp Val Tyr Cys Ser Asp Lys Thr Ser Ala Glu Trp
                85                  90                  95

Trp Asn Lys Asn Leu Tyr Pro Leu Ile Ile Lys Ser Ala Asn Asp Ile
            100                 105                 110

Ala Ser Tyr Gly Phe Lys Val Ala Gly Asp Pro Ser Ile Lys Lys Asp
        115                 120                 125

Gly Tyr Ph  Lys Lys Leu Gln Asp  lu Leu Asp Asn Ile Val Asp Asn
    130                 135                 140

Asn Ser Asp Asp Asp Ala Ile Ala Lys Ala Ile Lys Asp Phe Lys Ala
145                 150                 155                 160
```

Arg Cys ly Ile Leu Ile Lys Glu Ala Lys ln Tyr lu Glu Ala Ala
165 170 175

Lys Asn Ile Val Thr Ser Leu Asp Gln Phe Leu His Gly Asp Gln Lys
180 185 190

Lys Leu Glu Gly Val Ile Asn Ile Gln Lys Arg Leu Lys Glu Val Gln
195 200 205

Thr Ala Leu Asn Gln Ala His Gly Glu Ser Ser Pro Ala His Lys Glu
210 215 220

Leu Leu Glu Lys Val Lys Asn Leu Lys Thr Thr Leu Glu Arg Thr Ile
225 230 235 240

Lys Ala Glu Gln Asp Leu Glu Lys Lys Val Glu Tyr Ser Phe Leu Leu
245 250 255

Gly Pro Leu Leu Gly Phe Val Val Tyr Glu Ile Leu Glu Asn Thr Ala
260 265 270

Val Gln His Ile Lys Asn Gln Ile Asp Glu Ile Lys Lys Gln Leu Asp
275 280 285

Ser Ala Gln His Asp Leu Asp Arg Asp Val Lys Ile Ile Gly Met Leu
290 295 300

Asn Ser Ile Asn Thr Asp Ile Asp Asn Leu Tyr Ser Gln Gly Gln Glu
305 310 315 320

Ala Ile Lys Val Phe Gln Lys Leu Gln Gly Ile Trp Ala Thr Ile Gly
325 330 335

Ala Gln Ile Glu Asn Leu Arg Thr Thr Ser Leu Gln Glu Val Gln Asp
340 345 350

Ser Asp Asp Ala Asp Glu Ile Gln Ile Glu Leu Glu Asp Ala Ser Asp
355 360 365

Ala Trp Leu Val Val Ala Gln Glu Ala Arg Asp Phe Thr Leu Asn Ala
370 375 380

Tyr Ser Thr Asn Ser Arg Gln Asn Leu Pro Ile Asn Val Ile Ser Asp
385 390 395 400

Ser Cys Asn Cys Ser Thr Thr Asn Met Thr Ser Asn Gln Tyr Ser Asn
405 410 415

Pro Thr Thr Asn Met Thr Ser Asn Gln Tyr Met Ile Ser His Glu Tyr
420 425 430

Thr Ser Leu Pro Asn Asn Phe Met Leu Ser Arg Asn Ser Asn Leu Glu
435 440 445

Tyr Lys Cys Pro Glu Asn Asn Phe Met Ile Tyr Trp Tyr Asn Asn Ser
450 455 460

Asp Trp Tyr Asn Asn Ser Asp Trp Tyr Asn Asn
465 470 475

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1185 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: BACILLUS THURINGIENSIS
(C) INDIVIDUAL ISOLATE: PS69D1

(vii) IMMEDIATE SOURCE:
(B) CLONE: E. coli NM522(pMYC2317) NRRL B-18816

(ix) FEATURE:
(A) NAME/KEY: mat_peptide
(B) LOCATION: 1..1185

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

ATGATTTTAG GGAATGGAAA GACTTTACCA AAGCATATAA GATTAGCTCA TATTTTTGCA 60
ACACAGAATT CTTCAGCTAA GAAAGACAAT CCTCTTGGAC CAGAGGGGAT GGTTACTAAA 120
GACGGTTTTA TAATCTCTAA GGAAGAATGG GCATTTGTGC AGGCCTATGT GACTACAGGC 180
ACTGGTTTAC CTATCAATGA CGATGAGATG CGTAGACATG TTGGGTTACC ATCACGCATT 240
CAAATTCCTG ATGATTTTAA TCAATTATAT AAGGTTTATA ATGAAGATAA ACATTTATGC 300
AGTTGGTGGA ATGGTTTCTT GTTTCCATTA GTTCTTAAAA CAGCTAATGA TATTTCCGCT 360
TACGGATTTA AATGTGCTGG AAAGGGTGCC ACTAAAGGAT ATTATGAGGT CATGCAAGAC 420
GATGTAGAAA ATATTTCAGA TAATGGTTAT GATAAAGTTG CACAAGAAAA AGCACATAAG 480
GATCTGCAGG CGCGTTGTAA AATCCTTATT AAGGAGGCTG ATCAATATAA AGCTGCAGCG 540
GATGATGTTT CAAAACATTT AAACACATTT CTTAAAGGCG GTCAAGATTC AGATGGCAAT 600
GATGTTATTG GCGTAGAGGC TGTTCAAGTA CAACTAGCAC AAGTAAAAGA TAATCTTGAT 660
GGCCTATATG GCGACAAAAG CCCAAGACAT GAAGAGTTAC TAAAGAAAGT AGACGACCTG 720
AAAAAAGAGT TGGAAGCTGC TATTAAAGCA GAGAATGAAT TAGAAAAGAA AGTGAAAATG 780
AGTTTTGCTT TAGGACCATT ACTTGGATTT GTTGTATATG AAATCTTAGA GCTAACTGCG 840
GTCAAAAGTA TACACAAGAA AGTTGAGGCA CTACAAGCCG AGCTTGACAC TGCTAATGAT 900
GAACTCGACA GAGATGTAAA AATCTTAGGA ATGATGAATA GCATTGACAC TGATATTGAC 960
AACATGTTAG AGCAAGGTGA GCAAGCTCTT GTTGTATTTA GAAAAATTGC AGGCATTTGG 1020
AGTGTTATAA GTCTTAATAT CGGCAATCTT CGAGAAACAT CTTTAAAAGA GATAGAAGAA 1080
GAAAATGATG ACGATGCACT GTATATTGAG CTTGGTGATG CCGCTGGTCA ATGGAAAGAG 1140
ATAGCCGAGG AGGCACAATC CTTTGTACTA AATGCTTATA CTCCT 1185

(2) INFORMATION FOR SEQ ID NO:9:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 395 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: YES

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: BACILLUS THURINGIENSIS
(C) INDIVIDUAL ISOLATE: PS69D1

(vii) IMMEDIATE SOURCE:
(B) CLONE: E. coli NM522(pMYC2317) NRRL B-18816

(ix) FEATURE:
(A) NAME/KEY: Protein
(B) LOCATION: 1..395

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Met Ile Leu Gly Asn Gly Lys Thr Leu Pro Lys His Ile Arg Leu Ala
1 5 10 15

His Ile Phe Ala Thr Gln Asn Ser Ser Ala Lys Lys Asp Asn Pro Leu
20 25 30

Gly Pro Glu Gly Met Val Thr Lys Asp Gly Phe Ile Ile Ser Lys Glu
35 40 45

Glu Trp Ala Phe Val Gln Ala Tyr Val Thr Thr Gly Thr Gly Leu Pro
50 55 60

Ile Asn Asp Asp Glu Met Arg Arg His Val Gly Leu Pro Ser Arg Il
65 70 75 80

Gln Ile Pro Asp Asp Phe Asn Gln Leu Tyr Lys Val Tyr Asn lu Asp
85 90 95

Lys His Leu Cys Ser Trp Trp Asn Gly Phe Leu Phe Pro Leu Val Leu
100 105 110

Lys Thr Ala Asn Asp Ile Ser Ala Tyr Gly Phe Lys Cys Ala Gly Lys
115 120 125

Gly Ala Thr Lys Gly Tyr Tyr Glu Val Met Gln Asp Asp Val Glu Asn
130 135 140

Ile Ser Asp Asn Gly Tyr Asp Lys Val Ala Gln Glu Lys Ala His Lys
145 150 155 160

Asp Leu Gln Ala Arg Cys Lys Ile Leu Ile Lys Glu Ala Asp Gln Tyr
165 170 175

Lys Ala Ala Ala Asp Asp Val Ser Lys His Leu Asn Thr Phe Leu Lys
180 185 190

Gly Gly Gln Asp Ser Asp Gly Asn Asp Val Ile Gly Val Glu Ala Val
195 200 205

Gln Val Gln Leu Ala Gln Val Lys Asp Asn Leu Asp Gly Leu Tyr Gly
210 215 220

Asp Lys Ser Pro Arg His Glu Glu Leu Leu Lys Lys Val Asp Asp Leu
225 230 235 240

Lys Lys Glu Leu Glu Ala Ala Ile Lys Ala Glu Asn Glu Leu Glu Lys
245 250 255

Lys Val Lys Met Ser Phe Ala Leu Gly Pro Leu Leu Gly Phe Val Val
260 265 270

Tyr Glu Ile Leu Glu Leu Thr Ala Val Lys Ser Ile His Lys Lys Val
275 280 285

Glu Ala Leu Gln Ala Glu Leu Asp Thr Ala Asn Asp Glu Leu Asp Arg
290 295 300

Asp Val Lys Ile Leu Gly Met Met Asn Ser Ile Asp Thr Asp Ile Asp
305 310 315 320

Asn Met Leu Glu Gln Gly Glu Gln Ala Leu Val Val Phe Arg Lys Ile
325 330 335

Ala Gly Ile Trp Ser Val Ile Ser Leu Asn Ile Gly Asn Leu Arg Glu
340 345 350

Thr Ser Leu Lys Glu Ile Glu Glu Glu Asn Asp Asp Asp Ala Leu Tyr
355 360 365

Ile Glu Leu Gly Asp Ala Ala Gly Gln Trp Lys Glu Ile Ala Glu Glu
370 375 380

Ala Gln Ser Phe Val Leu Asn Ala Tyr Thr Pro
385 390 395

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 22 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

AGARTRKWTW AATGGWGCKM AW 22

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 8 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

Pro Thr Phe Asp Pro Asp Leu Tyr
1 5

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 14 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Ala Ile Leu Asn Glu Leu Tyr Pro Ser Val Pro Tyr Asn Val
1 5 10

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 14 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Ala Ile Leu Asn Glu Leu Tyr Pro Ser Val Pro Tyr Asn Val
1 5 10

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 17 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Ile Ile Asp Ser Lys Thr Thr Leu Pro Arg His Ser Leu Ile Asn
1 5 10 15

Thr

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 24 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Met Ile Leu Gly Asn Gly Lys Thr Leu Pro Lys His Ile Arg Leu Ala
1 5 10 15

His Ile Phe Ala Thr Gln Asn Ser
20

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

GCAATTTTAA ATGAATTATA TCC 23

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 56 bases
(B) TYPE: nucleic acid

(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

ATGATTATTG ATTCTAAAAC AACATTACCA AGACATTCWT TAATWAATAC WATWAA 56

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 38 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

AAACATATTA GATTAGCACA TATTTTTGCA ACACAAAA 38

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 17 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

CAAYTACAAG CWCAACC 17

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

AGGAACAAAY TCAAKWCGRT CTA 23

(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

TGGAATAAAT TCAATTYKRT CWA 23

(2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 28 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

TGATTTTWMT CAATTATATR AKGTTTAT 28

(2) INFORMATION FOR SEQ ID NO:23:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

AAGAGTTAYT ARARAAAGTA 20

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 35 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

TTAGGACCAT TRYTWGGATT TGTTGTWTAT GAAAT 35

(2) INFORMATION FOR SEQ ID NO:25:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 27 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

GAYAGAGATG TWAAAATYWT AGGAATG 27

(2) INFORMATION FOR SEQ ID NO:26:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (synthetic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

TTMTTAAAWC WGCTAATGAT ATT 23

(2) INFORMATION FOR SEQ ID NO:27:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1425 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: BACILLUS THURINGIENSIS
(C) INDIVIDUAL ISOLATE: PS86A1

(vii) IMMEDIATE SOURCE:
(B) CLONE: E. coli NM522(pMYC1638) NRRL B-18751

(ix) FEATURE:
(A) NAME/KEY: mat_peptide
(B) LOCATION: 1..1425

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

```
ATGATTATTG ATAGTAAAAC GACTTTACCT AGACATTCAC TTATTCATAC AATTAAATTA      60
AATTCTAATA AGAAATATGG TCCTGGTGAT ATGACTAATG GAAATCAATT TATTATTTCA     120
AAACAAGAAT GGGCTACGAT TGGAGCATAT ATTCAGACTG GATTAGGTTT ACCAGTAAAT     180
CAACAACAAT TAAGAACACA TGTTAATTTA AGTCAGGATA TATCAATACC TAGTGATTTT     240
TCTCAATTAT ATGATGTTTA TTGTTCTGAT AAAACTTCAG CAGAATGGTG GAATAAAAAT     300
TTATATCCTT TAATTATTAA ATCTGCTAAT GATATTGCTT CATATGGTTT TAAAGTTGCT     360
GGTGATCCTT CTATTAAGAA AGATGGATAT TTTAAAAAAT TGCAAGATGA ATTAGATAAT     420
ATTGTTGATA ATAATTCCGA TGATGATGCA ATAGCTAAAG CTATTAAAGA TTTTAAAGCG     480
CGATGTGGTA TTTTAATTAA AGAAGCTAAA CAATATGAAG AAGCTGCAAA AAATATTGTA     540
ACATCTTTAG ATCAATTTTT ACATGGTGAT CAGAAAAAAT TAGAAGGTGT TATCAATATT     600
CAAAAACGTT TAAAAGAAGT TCAAACAGCT CTTAATCAAG CCCATGGGGA AAGTAGTCCA     660
GCTCATAAAG AGTTATTAGA AAAAGTAAAA AATTTAAAAA CAACATTAGA AAGGACTATT     720
AAAGCTGAAC AAGATTTAGA GAAAAAAGTA GAATATAGTT TTCTATTAGG ACCATTGTTA     780
GGATTTGTTG TTTATGAAAT TCTTGAAAAT ACTGCTGTTC AGCATATAAA AAATCAAATT     840
GATGAGATAA AGAAACAATT AGATTCTGCT CAGCATGATT TGGATAGAGA TGTTAAAATT     900
ATAGGAATGT TAAATAGTAT TAATACAGAT ATTGATAATT TATATAGTCA AGGACAAGAA     960
GCAATTAAAG TTTTCCAAAA GTTACAAGGT ATTTGGGCTA CTATTGGAGC TCAAATAGAA    1020
AATCTTAGAA CAACGTCGTT ACAAGAAGTT CAAGATTCTG ATGATGCTGA TGAGATACAA    1080
ATTGAACTTG AGGACGCTTC TGATGCTTGG TTAGTTGTGG CTCAAGAAGC TCGTGATTTT    1140
ACACTAAATG CTTATTCAAC TAATAGTAGA CAAAATTTAC CGATTAATGT TATATCAGAT    1200
TCATGTAATT GTTCAACAAC AAATATGACA TCAAATCAAT ACAGTAATCC AACAACAAAT    1260
ATGACATCAA ATCAATATAT GATTTCACAT GAATATACAA GTTTACCAAA TAATTTTATG    1320
TTATCAAGAA ATAGTAATTT AGAATATAAA TGTCCTGAAA ATAATTTTAT GATATATTGG    1380
TATAATAATT CGGATTGGTA TAATAATTCG GATTGGTATA ATAAT                    1425
```

(2) INFORMATION FOR SEQ ID NO:28:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 475 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: YES

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: BACILLUS THURINGIENSIS
(C) INDIVIDUAL ISOLATE: PS86A1

(vii) IMMEDIATE SOURCE:
(B) CLONE: E. coli NM522(pMYC1638) NRRL B-18751

(ix) FEATURE:
(A) NAME/KEY: Protein
(B) LOCATION: 1..475

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:28:

```
Met Ile Ile Asp Ser Lys Thr Thr Leu Pro Arg His Ser Leu Ile His
1               5                  10                  15

Thr Ile Lys Leu Asn Ser Asn Lys Lys Tyr Gly Pro Gly Asp Met Thr
            20                  25                  30

Asn Gly Asn Gln Phe Ile Il  Ser Lys  ln Glu Trp Ala Thr Ile Gly
        35                  40                  45
```

Ala Tyr Ile ln Thr Gly Leu ly Leu Pro Val Asn Glu Gln Gln Leu
50 55 60

Arg Thr His Val Asn Leu Ser Gln Asp Ile Ser Ile Pro Ser Asp Phe
65 70 75 80

Ser Gln Leu Tyr Asp Val Tyr Cys Ser Asp Lys Thr Ser Ala Glu Trp
85 90 95

Trp Asn Lys Asn Leu Tyr Pro Leu Ile Ile Lys Ser Ala Asn Asp Ile
100 105 110

Ala Ser Tyr Gly Phe Lys Val Ala Gly Asp Pro Ser Ile Lys Lys Asp
115 120 125

Gly Tyr Phe Lys Lys Leu Gln Asp Glu Leu Asp Asn Ile Val Asp Asn
130 135 140

Asn Ser Asp Asp Asp Ala Ile Ala Lys Ala Ile Lys Asp Phe Lys Ala
145 150 155 160

Arg Cys Gly Ile Leu Ile Lys Glu Ala Lys Gln Tyr Glu Glu Ala Ala
165 170 175

Lys Asn Ile Val Thr Ser Leu Asp Gln Phe Leu His Gly Asp Gln Lys
180 185 190

Lys Leu Glu Gly Val Ile Asn Ile Gln Lys Arg Leu Lys Glu Val Gln
195 200 205

Thr Ala Leu Asn Gln Ala His Gly Glu Ser Ser Pro Ala His Lys Glu
210 215 220

Leu Leu Glu Lys Val Lys Asn Leu Lys Thr Thr Leu Glu Arg Thr Ile
225 230 235 240

Lys Ala Glu Gln Asp Leu Glu Lys Lys Val Glu Tyr Ser Phe Leu Leu
245 250 255

Gly Pro Leu Leu Gly Phe Val Val Tyr Glu Ile Leu Glu Asn Thr Ala
260 265 270

Val Gln His Ile Lys Asn Gln Ile Asp Glu Ile Lys Lys Gln Leu Asp
275 280 285

Ser Ala Gln His Asp Leu Asp Arg Asp Val Lys Ile Ile Gly Met Leu
290 295 300

Asn Ser Ile Asn Thr Asp Ile Asp Asn Leu Tyr Ser Gln Gly Gln Glu
305 310 315 320

Ala Ile Lys Val Phe Gln Lys Leu Gln Gly Ile Trp Ala Thr Ile Gly
325 330 335

Ala Gln Ile Glu Asn Leu Arg Thr Thr Ser Leu Gln Glu Val Gln Asp
340 345 350

Ser Asp Asp Ala Asp Glu Ile Gln Ile Glu Leu Glu Asp Ala Ser Asp
355 360 365

Ala Trp Leu Val Val Ala Gln Glu Ala Arg Asp Phe Thr Leu Asn Ala
370 375 380

Tyr Ser Thr Asn Ser Arg Gln Asn Leu Pro Ile Asn Val Ile Ser Asp
385 390 395 400

Ser Cys Asn Cys Ser Thr Thr Asn Met Thr Ser Asn Gln Tyr Ser Asn
405 410 415

Pro Thr Thr Asn Met Thr Ser Asn Gln Tyr Met Ile Ser His Glu Tyr
420 425 430

Thr Ser Leu Pro Asn Asn Phe Met Leu Ser Arg Asn Ser Asn Leu Glu
435 440 445

Tyr Lys Cys Pro Glu Asn Asn Phe Met Ile Tyr Trp Tyr Asn Asn Ser
450 455 460

Asp Trp Tyr Asn Asn Ser Asp Trp Tyr Asn Asn
465 470 475

(2) INFORMATION FOR SEQ ID NO:29:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 3471 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: Bacillus thuringiensis
(B) STRAIN: kumamotoensis
(C) INDIVIDUAL ISOLATE: PS50C

(vii) IMMEDIATE SOURCE:
(B) CLONE: E. coli NM522(pMYC2320) NRRL B-18769

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:29:

ATGAGTCCAA ATAATCAAAA TGAATATGAA ATTATAGATG CGACACCTTC TACATCTGTA 60
TCCAGTGATT CTAACAGATA CCCTTTTGCG AATGAGCCAA CAGATGCGTT ACAAAATATG 120
AATTATAAAG ATTATCTGAA AATGTCTGGG GGAGAGAATC CTGAATTATT TGGAAATCCG 180
GAGACGTTTA TTAGTTCATC CACGATTCAA ACTGGAATTG GCATTGTTGG TCGAATACTA 240
GGAGCTTTAG GGGTTCCATT TGCTAGTCAG ATAGCTAGTT TCTATAGTTT CATTGTTGGT 300
CAATTATGGC CGTCAAAGAG CGTAGATATA TGGGGAGAAA TTATGGAACG AGTGGAAGAA 360
CTCGTTGATC AAAAAATAGA AAAATATGTA AAAGATAAGG CTCTTGCTGA ATTAAAAGGG 420
CTAGGAAATG CTTTGGATGT ATATCAGCAG TCACTTGAAG ATTGGCTGGA AAATCGCAAT 480
GATGCAAGAA CTAGAAGTGT TGTTTCTAAT CAATTTATAG CTTTAGATCT TAACTTTGTT 540
AGTTCAATTC CATCTTTTGC AGTATCCGGA CACGAAGTAC TATTATTAGC AGTATATGCA 600
CAGGCTGTGA ACCTACATTT ATTGTTATTA AGAGATGCTT CTATTTTTGG AGAAGAGTGG 660
GGATTTACAC CAGGTGAAAT TTCTAGATTT TATAATCGTC AAGTGCAACT TACCGCTGAA 720
TATTCAGACT ATTGTGTAAA GTGGTATAAA ATCGGCTTAG ATAAATTGAA AGGTACCACT 780
TCTAAAAGTT GGCTGAATTA TCATCAGTTC CGTAGAGAGA TGACATTACT GGTATTAGAT 840
TTGGTGGCGT TATTTCCAAA CTATGACACA CATATGTATC CAATCGAAAC AACAGCTCAA 900
CTTACACGGG ATGTGTATAC AGATCCGATA GCATTTAACA TAGTGACAAG TACTGGATTC 960
TGCAACCCTT GGTCAACCCA CAGTGGTATT CTTTTTTATG AAGTTGAAAA CAACGTAATT 1020
CGTCCGCCAC ACTTGTTTGA TATACTCAGC TCAGTAGAAA TTAATACAAG TAGAGGGGGT 1080
ATTACGTTAA ATAATGATGC ATATATAAAC TACTGGTCAG GACATACCCT AAAATATCGT 1140
AGAACAGCTG ATTCGACCGT AACATACACA GCTAATTACG GTCGAATCAC TTCAGAAAAG 1200
AATTCATTTG CACTTGAGGA TAGGGATATT TTTGAAATTA ATTCAACTGT GGCAAACCTA 1260
GCTAATTACT ACCAAAAGGC ATATGGTGTG CCGGGATCTT GGTTCCATAT GGTAAAAAGG 1320
GGAACCTCAT CAACAACAGC GTATTTATAT TCAAAAACAC ATACAGCTCT CCAAGGGTGT 1380
ACACAGGTTT ATGAATCAAG TGATGAAATA CCTCTAGATA GAACTGTACC GGTAGCTGAA 1440
AGCTATAGTC ATAGATTATC TCATATTACC TCCCATTCTT TCTCTAAAAA TGGGAGTGCA 1500
TACTATGGGA GTTTCCCTGT ATTTGTTTGG ACACATACTA GTGCGGATTT AAATAATACA 1560
ATATATTCAG ATAAAATCAC TCAAATTCCA GCGGTAAAGG GAGACATGTT ATATCTAGGG 1620
GGTTCCGTAG TACAGGGTCC TGGATTTACA GGAGGAGATA TATTAAAAAG AACCAATCCT 1680
AGCATATTAG GGACCTTTGC GGTTACAGTA AATGGGTCGT TATCACAAAG ATATCGTGTA 1740
AGAATTCGCT ATGCCTCTAC AACAGATTTT GAATTTACTC TATACCTTGG CGACACAATA 1800
GAAAAAAATA GATTTAACAA AACTATGGAT AATGGGGCAT CTTTAACGTA TGAAACATTT 1860
AAATTCGCAA GTTTCATTAC TGATTTCCAA TTCAGAGAAA CACAAGATAA AATACTCCTA 1920
TCCATGGGTG ATTTTAGCTC CGGTCAAGAA GTTTATATAG ACCGAATCGA ATTCATCCCA 1980
GTAGATGAGA CATATGAGGC GGAACAAGAT TTAGAAGCGG CGAAGAAAGC AGTGAATGCC 2040
TTGTTTACGA ATACAAAAGA TGGCTTACGA CCAGGTGTAA CGGATTATGA AGTAAATCAA 2100

GCGGCAAACT TAGTGGAATG CCTATCGGAT GATTTATATC CAAATGAAAA ACGATTGTTA 2160
TTTGATGCGG TGAGAGAGGC AAAACGCCTC AGTGGGGCAC GTAACTTACT ACAAGATCCA 2220
GATTTCCAAG AGATAAACGG AGAAAATGGA TGGGCGGCAA GTACGGGAAT TGAGATTGTA 2280
GAAGGGGATG CTGTATTTAA AGGACGTTAT CTACGCCTAC CAGGTGCACG AGAAATTGAT 2340
ACGGAAACGT ATCCAACGTA TCTGTATCAA AAAGTAGAGG AAGGTGTATT AAAACCATAC 2400
ACAAGATATA GACTGAGAGG GTTTGTGGGA AGTAGTCAAG GATTAGAAAT TTATACGATA 2460
CGTCACCAAA CGAATCGAAT TGTAAAGAAT GTACCAGATG ATTTATTGCC AGATGTATCT 2520
CCTGTAAACT CTGATGGCAG TATCAATCGA TGCAGCGAAC AAAAGTATGT GAATAGCCGT 2580
TTAGAAGGAG AAAACCGTTC TGGTGATGCA CATGAGTTCT CGCTCCCTAT CGATATAGGA 2640
GAGCTGGATT ACAATGAAAA TGCAGGAATA TGGGTTGGAT TTAAGATTAC GGACCCAGAG 2700
GGATACGCAA CACTTGGAAA TCTTGAATTA GTCGAAGAGG GACCTTTGTC AGGAGACGCA 2760
TTAGAGCGCT TGCAAAGAGA AGAACAACAG TGGAAGATTC AAATGACAAG AAGACGTGAA 2820
GAGACAGATA GAAGATACAT GGCATCGAAA CAAGCGGTAG ATCGTTTATA TGCCGATTAT 2880
CAGGATCAAC AACTGAATCC TGATGTAGAG ATTACAGATC TTACTGCGGC TCAAGATCTG 2940
ATACAGTCCA TTCCTTACGT ATATAACGAA ATGTTCCCAG AAATACCAGG GATGAACTAT 3000
ACGAAGTTTA CAGAATTAAC AGATCGACTC CAACAAGCGT GGAATTTGTA TGATCAGCGA 3060
AATGCCATAC CAAATGGTGA TTTTCGAAAT GGGTTAAGTA ATTGGAATGC AACGCCTGGC 3120
GTAGAAGTAC AACAAATCAA TCATACATCT GTCCTTGTGA TTCCAAACTG GGATGAACAA 3180
GTTTCACAAC AGTTTACAGT TCAACCGAAT CAAAGATATG TATTACGAGT TACTGCAAGA 3240
AAAGAAGGGG TAGGAAATGG ATATGTAAGT ATTCGTGATG GTGGAAATCA ATCAGAAACG 3300
CTTACTTTTA GTGCAAGCGA TTATGATACA AATGGTGTGT ATAATGACCA AACCGGCTAT 3360
ATCACAAAAA CAGTGACATT CATCCCGTAT ACAGATCAAA TGTGGATTGA AATAAGTGAA 3420
ACAGAAGGTA CGTTCTATAT AGAAAGTGTA GAATTGATTG TAGACGTAGA G 3471

(2) INFORMATION FOR SEQ ID NO:30:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1157 amino acids
(B) TYPE: amino acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(iii) HYPOTHETICAL: YES

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: Bacillus thuringiensis
(B) STRAIN: kumamotoensis
(C) INDIVIDUAL ISOLATE: PS50C

(vii) IMMEDIATE SOURCE:
(B) CLONE: E. coli NM522(pMYC2320) NRRL B-18769

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:30:

Met Ser Pro Asn Asn Gln Asn Glu Tyr Glu Ile Ile Asp Ala Thr Pro
1 5 10 15

Ser Thr Ser Val Ser Ser Asp Ser Asn Arg Tyr Pro Phe Ala Asn Glu
20 25 30

Pro Thr Asp Ala Leu Gln Asn Met Asn Tyr Lys Asp Tyr Leu Lys Met
35 40 45

Ser Gly Gly Glu Asn Pro Glu Leu Phe Gly Asn Pro Glu Thr Phe Ile
50 55 60

Ser Ser Ser Thr Il Gln Thr ly Ile Gly Ile Val Gly Arg Ile Leu
65 70 75 80

Gly Ala Leu Gly Val Pro Phe Ala Ser Gln Ile Ala Ser Phe Tyr Ser
85 90 95

Phe Ile Val Gly Gln Leu Trp Pro Ser Lys Ser Val Asp Ile Trp Gly
100 105 110

Glu Ile Met Glu Arg Val Glu lu Leu Val Asp Gln Lys Ile lu Lys
115 120 125

Tyr Val Lys Asp Lys Ala Leu Ala Glu Leu Lys Gly Leu Gly Asn Ala
130 135 140

Leu Asp Val Tyr Gln Gln Ser Leu Glu Asp Trp Leu Glu Asn Arg Asn
145 150 155 160

Asp Ala Arg Thr Arg Ser Val Val Ser Asn Gln Phe Ile Ala Leu Asp
165 170 175

Leu Asn Phe Val Ser Ser Ile Pro Ser Phe Ala Val Ser Gly His Glu
180 185 190

Val Leu Leu Leu Ala Val Tyr Ala Gln Ala Val Asn Leu His Leu Leu
195 200 205

Leu Leu Arg Asp Ala Ser Ile Phe Gly Glu Glu Trp Gly Phe Thr Pro
210 215 220

Gly Glu Ile Ser Arg Phe Tyr Asn Arg Gln Val Gln Leu Thr Ala Glu
225 230 235 240

Tyr Ser Asp Tyr Cys Val Lys Trp Tyr Lys Ile Gly Leu Asp Lys Leu
245 250 255

Lys Gly Thr Thr Ser Lys Ser Trp Leu Asn Tyr His Gln Phe Arg Arg
260 265 270

Glu Met Thr Leu Leu Val Leu Asp Leu Val Ala Leu Phe Pro Asn Tyr
275 280 285

Asp Thr His Met Tyr Pro Ile Glu Thr Thr Ala Gln Leu Thr Arg Asp
290 295 300

Val Tyr Thr Asp Pro Ile Ala Phe Asn Ile Val Thr Ser Thr Gly Phe
305 310 315 320

Cys Asn Pro Trp Ser Thr His Ser Gly Ile Leu Phe Tyr Glu Val Glu
325 330 335

Asn Asn Val Ile Arg Pro Pro His Leu Phe Asp Ile Leu Ser Ser Val
340 345 350

Glu Ile Asn Thr Ser Arg Gly Gly Ile Thr Leu Asn Asn Asp Ala Tyr
355 360 365

Ile Asn Tyr Trp Ser Gly His Thr Leu Lys Tyr Arg Arg Thr Ala Asp
370 375 380

Ser Thr Val Thr Tyr Thr Ala Asn Tyr Gly Arg Ile Thr Ser Glu Lys
385 390 395 400

Asn Ser Phe Ala Leu Glu Asp Arg Asp Ile Phe Glu Ile Asn Ser Thr
405 410 415

Val Ala Asn Leu Ala Asn Tyr Tyr Gln Lys Ala Tyr Gly Val Pro Gly
420 425 430

Ser Trp Phe His Met Val Lys Arg Gly Thr Ser Ser Thr Thr Ala Tyr
435 440 445

Leu Tyr Ser Lys Thr His Thr Ala Leu Gln Gly Cys Thr Gln Val Tyr
450 455 460

Glu Ser Ser Asp Glu Ile Pro Leu Asp Arg Thr Val Pro Val Ala Glu
465 470 475 480

Ser Tyr Ser His Arg Leu Ser His Ile Thr Ser His Ser Phe Ser Lys
485 490 495

Asn Gly Ser Ala Tyr Tyr Gly Ser Phe Pro Val Phe Val Trp Thr His
500 505 510

Thr Ser Ala Asp Leu Asn Asn Thr Ile Tyr Ser Asp Lys Ile Thr Gln
515 520 525

Ile Pro Ala Val Lys Gly Asp Met Leu Tyr Leu Gly Gly Ser Val Val
530 535 540

Gln Gly Pro Gly Phe Thr Gly Gly Asp Ile Leu Lys Arg Thr Asn Pro
545 550 555 560

Ser Ile Leu Gly Thr Ph Ala Val Thr Val Asn Gly Ser Leu Ser Gln
565 570 575

Arg Tyr Arg Val Arg Ile Arg Tyr Ala Ser Thr Thr Asp Phe Glu Phe
580 585 590

Thr Leu Tyr Leu Gly Asp Thr Ile Glu Lys Asn Arg Phe Asn Lys Thr
595 600 605

Met Asp Asn Gly Ala Ser Leu Thr Tyr Glu Thr Phe Lys Phe Ala Ser
610 615 620

Phe Ile Thr Asp Phe Gln Phe Arg Glu Thr Gln Asp Lys Ile Leu Leu
625 630 635 640

Ser Met Gly Asp Phe Ser Ser Gly Gln Glu Val Tyr Ile Asp Arg Ile
645 650 655

Glu Phe Ile Pro Val Asp Glu Thr Tyr Glu Ala Glu Gln Asp Leu Glu
660 665 670

Ala Ala Lys Lys Ala Val Asn Ala Leu Phe Thr Asn Thr Lys Asp Gly
675 680 685

Leu Arg Pro Gly Val Thr Asp Tyr Glu Val Asn Gln Ala Ala Asn Leu
690 695 700

Val Glu Cys Leu Ser Asp Asp Leu Tyr Pro Asn Glu Lys Arg Leu Leu
705 710 715 720

Phe Asp Ala Val Arg Glu Ala Lys Arg Leu Ser Gly Ala Arg Asn Leu
725 730 735

Leu Gln Asp Pro Asp Phe Gln Glu Ile Asn Gly Glu Asn Gly Trp Ala
740 745 750

Ala Ser Thr Gly Ile Glu Ile Val Glu Gly Asp Ala Val Phe Lys Gly
755 760 765

Arg Tyr Leu Arg Leu Pro Gly Ala Arg Glu Ile Asp Thr Glu Thr Tyr
770 775 780

Pro Thr Tyr Leu Tyr Gln Lys Val Glu Glu Gly Val Leu Lys Pro Tyr
785 790 795 800

Thr Arg Tyr Arg Leu Arg Gly Phe Val Gly Ser Ser Gln Gly Leu Glu
805 810 815

Ile Tyr Thr Ile Arg His Gln Thr Asn Arg Ile Val Lys Asn Val Pro
820 825 830

Asp Asp Leu Leu Pro Asp Val Ser Pro Val Asn Ser Asp Gly Ser Ile
835 840 845

Asn Arg Cys Ser Glu Gln Lys Tyr Val Asn Ser Arg Leu Glu Gly Glu
850 855 860

Asn Arg Ser Gly Asp Ala His Glu Phe Ser Leu Pro Ile Asp Ile Gly
865 870 875 880

Glu Leu Asp Tyr Asn Glu Asn Ala Gly Ile Trp Val Gly Phe Lys Ile
885 890 895

Thr Asp Pro Glu Gly Tyr Ala Thr Leu Gly Asn Leu Glu Leu Val Glu
900 905 910

Glu Gly Pro Leu Ser Gly Asp Ala Leu Glu Arg Leu Gln Arg Glu Glu
915 920 925

Gln Gln Trp Lys Ile Gln Met Thr Arg Arg Arg Glu Glu Thr Asp Arg
930 935 940

Arg Tyr Met Ala Ser Lys Gln Ala Val Asp Arg Leu Tyr Ala Asp Tyr
945 950 955 960

Gln Asp Gln Gln Leu Asn Pro Asp Val Glu Ile Thr Asp Leu Thr Ala
965 970 975

Ala Gln Asp Leu Ile Gln Ser Ile Pro Tyr Val Tyr Asn Glu Met Phe
980 985 990

Pro lu Ile Pro Gly Met Asn Tyr Thr Lys Phe Thr Glu Leu Thr Asp
995 1000 1005

Arg Leu Gln ln Ala Trp Asn Leu Tyr Asp ln Arg Asn Ala Ile Pro
1010 1015 1020

Asn Gly Asp Phe Arg Asn Gly Leu Ser Asn Trp Asn Ala Thr Pro Gly
1025 1030 1035 1040

Val lu Val Gln Gln Ile Asn His Thr Ser Val Leu Val Ile Pro Asn
1045 1050 1055

Trp Asp Glu Gln Val Ser Gln Gln Phe Thr Val Gln Pro Asn Gln Arg
1060 1065 1070

Tyr Val Leu Arg Val Thr Ala Arg Lys Glu Gly Val Gly Asn Gly Tyr
1075 1080 1085

Val Ser Ile Arg Asp Gly Gly Asn Gln Ser Glu Thr Leu Thr Phe Ser
1090 1095 1100

Ala Ser Asp Tyr Asp Thr Asn Gly Val Tyr Asn Asp Gln Thr Gly Tyr
1105 1110 1115 1120

Ile Thr Lys Thr Val Thr Phe Ile Pro Tyr Thr Asp Gln Met Trp Ile
1125 1130 1135

Glu Ile Ser Glu Thr Glu Gly Thr Phe Tyr Ile Glu Ser Val Glu Leu
1140 1145 1150

Ile Val Asp Val Glu
1155

Claims

1. A method for controlling acarid pests wherein said method comprises contacting said pests with an acarid-controlling effective amount of a B.t. endotoxin.

2. The method, according to claim 1, wherein said toxin is obtainable from a B. t. isolate selected from the group consisting of B.t. PS50C, B.t. PS86A1, B.t. PS69D1, B.t. PS72L1, B.t. PS75J1, B.t. PS83E5, B.t. PS45B1, B.t. PS24J, B.t. PS94R3, B.t. PS17, B.t. PS62B1 and B.t. PS74G1, and mutants thereof.

3. The method, according to claim 2, wherein said isolate is PS50C.

4. The method, according to claim 2, wherein said isolate is PS86A1.

5. The method, according to claim 2, wherein said isolate is PS69D1.

6. The method, according to claim 2, wherein said isolate is PS72L2.

7. The method, according to claim 2, wherein said isolate is PS75J2.

8. The method, according to claim 2, wherein said isolate is PS83E5.

9. The method, according to claim 2, wherein said microbe is PS45B1.

10. The method, according to claim 2, wherein said isolate is PS24J.

11. The method, according to claim 2, wherein said isolate is PS94R3.

12. The method, according to claim 2, wherein said isolate is PS17.

13. The method, according to claim 2, wherein said isolate is PS62B1.

14. The method, according to claim 2, wherein said isolate is PS74G1.

15. The method, according to claim 3, wherein said toxin has the amino acid sequence of SEQ ID NO. 28.

16. The method, according to claim 4, wherein said toxin has the amino acid sequence of SEQ ID NO. 30.

17. The method, according to claim 5, wherein said toxin has the amino acid sequence of SEQ ID NO. 10.

18. The method, according to claim 12, wherein said toxin has the amino acid sequence of SEQ ID NO. 2.

19. The method, according to claim 12, wherein said toxin has the amino acid sequence of SEQ ID NO. 4.

20. The method, according to claim 1, wherein said acarid pest is a mite.

21. The method, according to claim 20, wherein said mite is the Two Spotted Spider Mite.

22. A composition of matter comprising a Bacillus thuringiensis isolate selected from the group consisting of B.t. PS72L1, B.t. PS75J1, B.t. PS83E5, B.t. PS45B1, B.t. PS24J, B.t. PS94R3, B.t. PS62B1 and B.t. PS74G1, and mutants thereof, or proteins, toxic crystals, or spores of said isolates, in association with an inert carrier.

23. The composition of matter, according to claim 22, comprising Bacillus thuringiensis PS24J.

24. The composition of matter, according to claim 22, comprising Bacillus thuringiensis PS94R3.

25. The composition of matter, according to claim 22, comprising Bacillus thuringiensis PS45B1.

26. The composition of matter, according to claim 22, comprising Bacillus thuringiensis PS62B1.

27. The composition of matter, according to claim 22, comprising Bacillus thuringiensis PS74G1.

28. The composition of matter, according to claim 22, comprising Bacillus thuringiensis PS72L1.

29. The composition of matter, according to claim 22, comprising Bacillus thuringiensis PS75J1.

30. The composition of matter, according to claim 22, comprising Bacillus thuringiensis PS83E5.

31. A composition for controlling an acaride pest wherein said composition comprises substantially intact, treated cells having pesticidal activity and prolonged persistence in the feeding zone of said pests when applied to the environment of acaride pests, wherein said pesticide is a polypeptide toxic to acaride pests, is intracellular, and is produced by a Bacillus thuringiensis isolate selected from the group consisting of B.t. PS50C, B.t. PS86A1, B.t. PS69D1, B.t. PS72L1, B.t. PS75J1, B.t. PS83E5, B.t. PS45B1, B.t. PS24J, B.t. PS94R3, B.t. PS17, B.t. PS62B1 and B.t. PS74G1, and mutants thereof.

32. The pesticidal composition, according to claim 18, wherein said treated cells are treated by chemical or physical means to prolong the pesticidal activity in the environment.

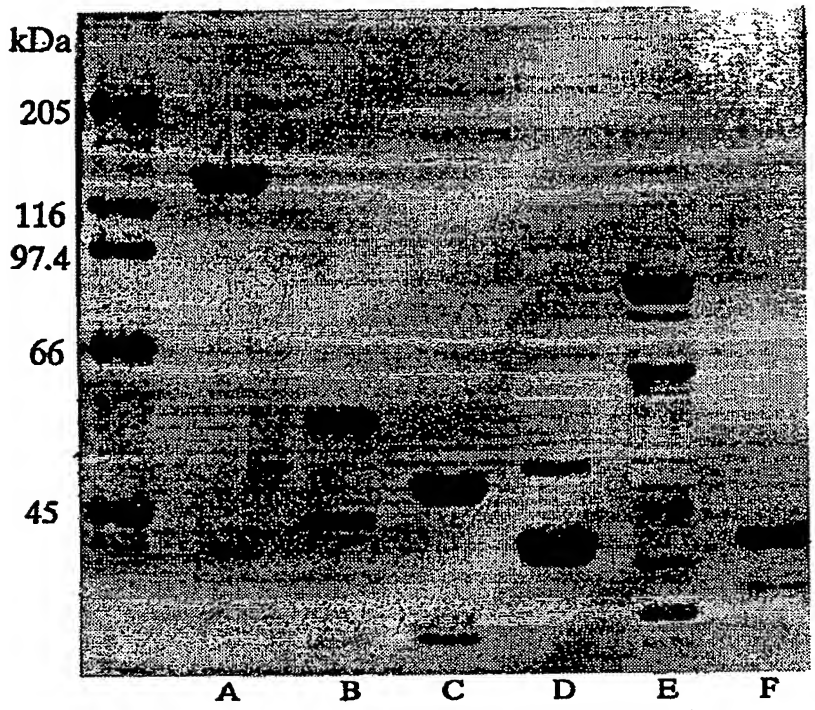
33. A gene encoding a toxin which is active against acarides wherein said gene is obtainable from a Bacillus thuringiensis isolate selected from the group consisting of B.t. PS72L1, B.t. PS75J1, B.t. PS83E5, B.t. PS45B1, B.t. PS24J, B.t. PS94R3, B.t. PS62B1 and B.t. PS74G1, and mutants thereof or is equivalent to one of said genes.

34. A toxin encoded by a gene obtainable from a Bacillus thuringiensis isolate selected from the group consisting of B.t. PS72L1, B.t. PS75J1, B.t. PS83E5, B.t. PS45B1, B.t. PS24J, B.t. PS94R3, B.t. PS62B1 and B.t. PS74G1, and mutants thereof, wherein said toxin is active against acaride pests.

35. A transformed host selected from the group consisting of a plant, a microbe, and a baculovirus transformed by a gene obtainable from a Bacillus thuringiensis isolate selected from the group consisting of B.t. PS72L1, B.t. PS75J1, B.t. PS83E5, B.t. PS45B1, B.t. PS24J, B.t. PS94R3, B.t. PS62B1 and B.t. PS74G1, and mutants thereof.

36. A biologically pure culture of a Bacillus thuringiensis selected from the group consisting of B.t. PS72L1, B.t. PS75J1, B.t. PS83E5, B.t. PS45B1, B.t. PS24J, B.t. PS94R3, B.t. PS62B1 and B.t. PS74G1, and mutants thereof.

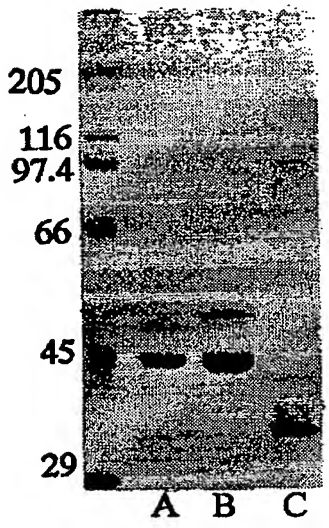
Figure 1



A. *Bacillus thuringiensis* PS50C
B. *Bacillus thuringiensis* PS86A1
C. *Bacillus thuringiensis* PS69D1
D. *Bacillus thuringiensis* PS72L1
E. *Bacillus thuringiensis* PS75J1
F. *Bacillus thuringiensis* PS83E5

2/3

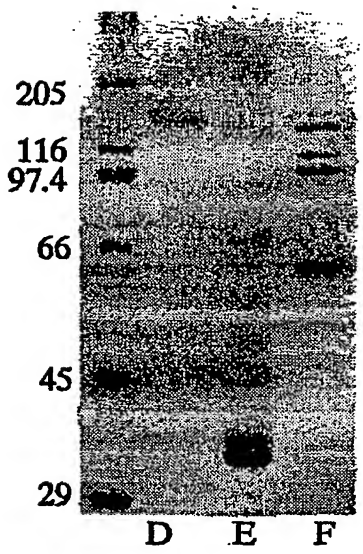
Figure 2A



A. Bacillus thuringiensis PS24J
B. Bacillus thuringiensis PS94R3
C. Bacillus thuringiensis PS45B1

3/3

Figure 2B



D. Bacillus thuringiensis PS17
E. Bacillus thuringiensis PS62B1
F. Bacillus thuringiensis PS74C1

INTERNATIONAL SEARCH REPORT

International Application No PCT/US 92/03546

I. CLASSIFICATION OF SUBJECT MATTER (if several classification symbols apply, indicate all)[6]

According to International Patent Classification (IPC) or to both National Classification and IPC

Int.Cl. 5 A01N63/00; A01N63/02; C12N1/20; //(C12N1/20, C12R1:07)

II. FIELDS SEARCHED

Minimum Documentation Searched[7]

Classification System	Classification Symbols
Int.Cl. 5	A01N ; C12R

Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched[8]

III. DOCUMENTS CONSIDERED TO BE RELEVANT[9]

Category[10]	Citation of Document,[11] with indication, where appropriate, of the relevant passages[12]	Relevant to Claim No.[13]
X	US,A,4 849 217 (G.G.SOARES ET. AL.) 18 July 1989 cited in the application see column 2, line 28 - column 3, line 40 see column 4, line 6 - column 5, line 25; claims	22, 25-27, 31,32,36
A	EP,A,0 303 426 (MYCOGEN CORPORATION) 15 February 1989	
A	JOURNAL OF ECONOMIC ENTOMOLOGY. vol. 83, no. 3, 1990, COLLEGE PARK, MARYLAND US pages 792 - 798; R.N.ROYALTY ET. AL.: 'Effects of Thuringiensin on Tetranychus urticae (Acari: Tetranychidae) Mortality, Fecundity, and Feeding.' cited in the application	

-/--

[10] Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier document but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

IV. CERTIFICATION

Date of the Actual Completion of the International Search	Date of Mailing of this International Search Report
16 SEPTEMBER 1992	25.09.92
International Searching Authority	**Signature of Authorized Officer**
EUROPEAN PATENT OFFICE	DONOVAN T.M.

Form PCT/ISA/210 (second sheet) (January 1985)

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)

Category *	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.
A	CHEMICAL ABSTRACTS, vol. 101, no. 23, 3 December 1984, Columbus, Ohio, US; abstract no. 206066P, R.A.LI ET. AL.: 'Biology of ectoparasites of birds and acaricides to control them.' page 222 ; see abstract & VET. ENTOMOL. AKAROL. 1983, USSR pages 263 - 272;	
A	JOURNAL OF INVERTEBRATE PATHOLOGY vol. 56, no. 2, September 1990, pages 258 - 266; S.C.MACINTOSH ET. AL.: 'Specificity and Efficacity of Purified Bacillus thuringiensis Proteins against Agronomically Important Insects'	

ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO. US 9203546 SA 60534

This annex lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on
The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information. 16/09/92

Patent document cited in search report	Publication date	Patent family member(s)		Publication date
US-A-4849217	18-07-89	None		
EP-A-0303426	15-02-89	US-A-	4948734	14-08-90
		JP-A-	1067192	13-03-89
		US-A-	5093120	03-03-92

EPO FORM P0479

For more details about this annex : see Official Journal of the European Patent Office, No. 12/82